

Original Article

Quantitative Analysis of Nucleic Acid Content in Spikevax (Moderna) and BNT162b2 (Pfizer) COVID-19 Vaccine Lots

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Abstract

Background: The emergence of mRNA-based COVID-19 vaccines represented a significant advancement in public health response during the pandemic. However, questions have arisen regarding the consistency of their nucleic acid content, the presence of undeclared genomic material, and the continued use of outdated viral sequences.

Methods: In this study, 17 lots of Spikevax (Moderna) and 7 lots of BNT162b2 (Pfizer) vaccines were analyzed for nucleic acid content using multiplex quantitative Real-Time PCR. Samples were assessed for mRNA identity, quantity, homogeneity, and the presence of undeclared nucleic acids, including DNA elements. The stability of nucleic acid content in expired vaccine lots stored at -80°C was also examined.

Results: Quantitative analysis confirmed the presence of mRNA sequences consistent with the Spikevax and BNT162b2 vaccines. However, variations in nucleic acid quantity were observed across lots. DNA sequences, including *Escherichia coli* genomic fragments, were detected in some samples. Despite the evolution of circulating SARS-CoV-2 variants, both vaccine types retained the original "Wuhan" S protein sequence. Expired vaccine samples exhibited reduced nucleic acid integrity. No evidence of SV40 was identified.

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Conclusion: The presence of undeclared DNA sequences and variability in nucleic acid content across lots underscores the need for enhanced quality control in vaccine manufacturing. The presence also raises InflammoThrombotic Immunologic Response (ITIR) concerns. Regulatory oversight should address the potential risks associated with genetic material inconsistencies to ensure vaccine safety and efficacy.

Keywords: COVID-19 vaccine; mRNA; Nucleic acid

Introduction

Infectious processes, including that caused by Severe Acute Respiratory Syndrome Coronavirus number 2 (SARS-CoV-2) result in an InflammoThrombotic Immunologic Response (ITIR) Disease (ITIRD) [1,2], in this instance Coronavirus Disease first described in 2019 (COVID-19). While medical management is possible [3], enhancement of immunity can be acquired through the administration of vaccines [4,5].

The development of mRNA-based COVID-19 vaccines, such as Spikevax (Moderna) and BNT162b2 (Pfizer), has been pivotal in mitigating the global pandemic. These vaccines rely on modified messenger RNA (mRNA) molecules to instruct cells to produce the SARS-CoV-2 spike (S) protein, triggering an immune response.

While mRNA vaccines underwent rigorous clinical evaluation, emerging concerns have prompted further examination of their nucleic acid content and batch consistency. In particular, discrepancies in genomic material, including the presence of undeclared DNA sequences, have raised questions about manufacturing integrity and potential biological risks.

This study was conducted in response to a formal request by Peter Kotlár, MD, Plenipotentiary for the Government of the Slovak Republic, to assess the nucleic acid content in multiple lots of Spikevax and BNT162b2 vaccines. The analysis aimed to:

- Identify the nucleic acids present in the vaccine preparations.
- Quantify nucleic acid content across multiple lots.
- Evaluate batch homogeneity.
- Determine whether undeclared genetic material was present.
- Assess potential biological implications of unexpected nucleic acids.
- Examine the effects of expired storage conditions on nucleic acid stability.
- The findings provide insights into vaccine composition, stability, and manufacturing practices, with implications for public health policy and regulatory oversight.

Methods

Terminology

Given the relative infancy of this field of genetic vaccines, we provide the following terminology.

mRNA – messenger RNA; a nucleic acid to convey the genetic information contained in it into a protein by translation, DNA – deoxyribonucleic acid, in this project used in the sense of double-stranded DNA (dsDNA), Multiplex quantitative Real-Time PCR – Real-time PCR, allowing quantitative simultaneous detection of several molecular targets at the same time, Expression cloning vector – a circular double-stranded DNA molecule, which is used for subcloning, propagation and expression of a transgene. Transgene – a foreign, often artificially created, synthetic nucleic acid construct (typically DNA) intended for the purposes of genetic modification of a recipient cell or organism.

GMO – genetically modified organism, and S protein – one of the SARS-CoV-2 proteins composing the viral spike which attaches to the surface of cells, the genetic sequence of which differs between individual variant strains of the virus.

Sample collection and handling

Twenty-four distinct vaccine lots (17 Spikevax, 7 BNT162b2) were provided under controlled conditions. Each lot contained 10 original, unopened vials stored at -80°C. Transport conditions were verified using an onboard thermometer to ensure continuous temperature control.

The package contained the following lots

Spikevax (Moderna): MV1013A, 200023A, 200156A, 223049, 200090A, 200106A, 200100A, 3005885, 3005836, 000090A, 000058A, 3005241, 3005697, 3006272, MV1018A, 400012A, 400011A.

BNT162b2 (Pfizer): FP9632, 1F1051A, 1LO84A, 1F1047A, 1F1059A, 1F1055A, PCB0020.

Nucleic acid isolation

Samples were thawed rapidly and processed using the QIAamp DNA Mini Kit (Qiagen, DE) following the manufacturer's protocol. DNA and RNA were eluted in 50 µl of elution buffer. Reverse transcription was performed using the verso cDNA Kit (ThermoFisher Scientific, USA) at 47°C for 1 hour. All isolates were subsequently stored at -80°C.

Analytical procedure used

From each tested lot (containing 10 individual vials in labeled boxes), 5 original, unopened, unused vials of the product were used. The remaining unused vials were left intact as reference material at -80°C. The residues after removing the appropriate volume for analysis (see below) were secured with parafilm and returned to -80°C for further reference.

DNA and RNA isolation

DNA and RNA isolation was performed using a commercial isolation kit QIAamp DNA Mini Kit (Qiagen, DE) according to the manufacturer's instructions. Vials of individual lots intended for processing were removed from -80°C and quickly thawed in a stream of air at ambient temperature. 500 µl of each sample was pipetted into pre-prepared tubes with Proteinase K and Lysis Buffer. After incubation for 10 minutes at 60°C, the lysate was precipitated with 96°C ethanol

and centrifuged through isolation columns. Isolation columns were then washed with buffers with different ethanol contents and the filters were air-dried. DNA and RNA were eluted into 50 µl of Elution Buffer.

4 µl of the DNA/RNA isolate each was used for reverse transcription using the verso cDNA kit (ThermoFisher Scientific, USA) according to the manufacturer's instructions. Reverse transcription was performed at 47°C for 1 hour.

DNA/RNA isolates were stored at -80°C, cDNA at -20°C.

PCR analysis

Multiplex quantitative Real-Time PCR was performed using oligonucleotides specific to:

- mRNA for the S protein (manufacturer-declared molecular target)
- DNA Ori sequence (contained in expression cloning vectors)
- DNA ITS *Escherichia coli* (to detect possible contamination from bacterial DNA used in vector propagation)

Molecular targets for quantitative multiplex Real-Time PCR

The following manufacturer-declared and manufacturer-non-declared molecular targets were used to analyze the content of individual lots at the nucleic acid level:

- mRNA for S protein, manufacturer-declared molecular target; sequence in Spikevax GenBank: OK120841; sequence in BNT162b2 GenBank: OR134577
- DNA Ori contained in the expression cloning vector, GenBank: OR134577
- DNA ITS *Escherichia coli* – Internal Transcribed Spacer in the 16S rDNA cassette, GenBank: AP027563, to assess the level of possible contamination of the samples with genomic DNA of *Escherichia coli* used for the propagation of expression vectors during the manufacturing processes.

Genomic reference sequences were obtained from GenBank, including

- Original Wuhan SARS-CoV-2 reference strain sequence (GenBank: MT192773) [6]
- Spikevax mRNA sequence (GenBank: OK120841) [7]
- BNT162b2 mRNA sequence (GenBank: OR134577) [8]

Results

Nucleic acid identification

PCR analysis confirmed the presence of mRNA sequences corresponding to the declared Spikevax and BNT162b2 vaccine profiles. Sequence alignments demonstrated significant differences between the vaccine mRNA sequences and the original Wuhan SARS-CoV-2 spike (S) protein mRNA sequence (GenBank: MT192773). These differences reflect intentional modifications made to enhance mRNA stability and translation efficiency. Despite these engineered differences, the encoded S protein in both vaccines contained only two amino acid substitutions (K986P and V987P) relative to the Wuhan strain. This stabilization strategy is consistent with manufacturer documentation.

Query	94	AGCCAGTGGGTGAACCTGACCACCCGGACCCAGCTGCCACCAGCCTACACCAACAGCTTC	153
Sbjct	36	AGTCAGTGTGTTAATCTTACAACCAGAACTCAATTACCCCTGCATACACTAATTCTTTC	95
Query	154	ACCCGGGGCGTCTACTACCCGACAAGGTGTTCCGGAGCAGCGTCCTGCACAGCACCAG	213
Sbjct	96	ACACGTGGTGTATTATTACCCTGACAAAGTTTTTCAGATCCTCAGTTTTACATTCAACTCAG	155
Query	214	GACCTGTTCTGCCCCTTCTTCAGCAACGTGACCTGGTTCACGCCATCCAGTGAGCGGC	273
Sbjct	156	GACTTGTCTTACCTTTCTTTTCCAATGTTACTTGGTTCATGCTATACATGTCTCTGGG	215
Query	274	ACCAACGGCACCAGCGGTTTCGACAACCCCGTGCTGCCCTTCAACGACGGCGTGTACTTC	333
Sbjct	216	ACCAATGGTACTAAGAGGTTTGATAACCCCTGTCCTACCATTAAATGATGGTGTATTATTT	275
Query	334	GCCAGCACCAGAGAAGAGCAACATCATCCGGGGCTGGATCTTCGGCACCACCCTGGACAGC	393
Sbjct	276	GCTTCCACTGAGAAGTCTAACATAATAAGAGGCTGGATTTTTGGTACTACTTTAGATTTCG	335
Query	394	AAGACCCAGAGCCTGCTGATCGTGAATAACGCCACCAACGTGGTGATCAAGGTGTGCGAG	453
Sbjct	336	AAGACCCAGTCCCTACTTATTGTTAATAACGCTACTAATGTGTATTAAAGTCTGTGAA	395
Query	454	TTCCAGTTCTGCAACGACCCCTTCTGGGGCTGTACTACCACAAGAACAACAAGAGCTGG	513
Sbjct	396	TTTCAATTTTGTAATGATCCATTTTTGGGTGTTTATTACCACAAAAACAACAAAAGTTGG	455
Query	514	ATGGAGAGCGAGTTCCGGGTGTACAGCAGCGCCAACAACCTGCACCTTCGAGTACGTGAGC	573
Sbjct	456	ATGGAAAGTGAGTTCAGAGTTTATTCTAGTGCGAATAATTGCACTTTTGAATATGTCTCT	515
Query	574	CAGCCCTTCTGATGGACCTGGAGGGCAAGCAGGGCAACTTCAAGAACCTGCGGGAGTTC	633
Sbjct	516	CAGCCTTTTCTTATGGACCTTGAAGGAAAAACAGGGTAATTTCAAAAATCTTAGGGAATTT	575
Query	634	GTGTTCAAGAACATCGACGGCTACTTCAAGATCTACAGCAAGCACACCCCAATCAACCTG	693
Sbjct	576	GTGTTTAAGAATATTGATGGTTATTTTAAATATATTCTAAGCACACGCCTATTAAATTA	635
Query	694	GTGCGGGATCTGCCCCAGGGCTTCTCAGCCCTGGAGCCCTGGTGGACCTGCCCATCGGC	753
Sbjct	636	GTGCGTGATCTCCCTCAGGGTTTTTCGGCTTTAGAACCATTGGTAGATTGCCAATAGGT	695
Query	754	ATCAACATCACCCGGTTCAGACCTGCTGGCCCTGCACCGGAGCTACCTGACCCAGGC	813
Sbjct	696	ATTAACATCACTAGGTTTCAAACCTTACTTGCTTTACATAGAAGTTATTGACTCCTGGT	755
Query	814	GA--CAGCAGCAGCGGGTGGACAGCAGGCGCGGCTGCTTACTACGTGGGCTACCTGCAGC	871
Sbjct	756	GATTCTTCTTCAG--GTTGGACAGCTGGTGTGTCAGCTTATTATGTGGGTATCTTCAAC	813
Query	872	CCCGGACCTTCTGCTGAAGTACAACGAGAACGGCACCATCACCGACGCCGTGGACTGCG	931
Sbjct	814	CTAGGACTTTTCTATTAAATATAATGAAATGGAACCATACAGATGCTGTAGACTGTG	873
Query	932	CCCTGGACCCTCTGAGCGAGACCAAGTGCACCCTGAAGAGCTTCACCGTGGAGAAGGGCA	991
Sbjct	874	CACCTTGACCCTCTCTCAGAAACAAAGTGACGTTGAAATCCTTCACTGTAGAAAAAGGAA	933
Query	992	TCTACCAGACCAGCAACTTCCGGGTGCAGCCACCGAGAGCATCGTGCGGTTCCCAACA	1051
Sbjct	934	TCTATCAAACCTTCTAACTTTAGAGTCCAACCAACAGAATCTATTGTTAGATTTCTAATA	993
Query	1052	TCACCAACCTGTGCCCTTTCGGCGAGGTGTTCAACGCCACCCGGTTTCGCCAGCGTGACG	1111
Sbjct	994	TTACAAACTTGTGCCCTTTTGGTGAAGTTTTTAACGCCACCAGATTGTCATCTGTTTATG	1053

Query	1112	CCTGGAACCGGAAGCGGATCAGCAACTGCGTGGCCGACTACAGCGTGCTGTACAACAGCG	1171
Sbjct	1054	CTTGGAACAGGAAGAGAATCAGCAACTGTGTTGCTGATTATTCTGTCCTATATAATTCCG	1113
Query	1172	C--CAGCTTCAGCACCTTCAAGTGCTACGGCGTGAGCCCCACCAAGCTGAACGACCTGTG	1229
Sbjct	1114	CATCATTTTC--CACTTTTAAGTGTTATGGAGTGTCTCCTACTAAATTAAATGATCTCTG	1171
Query	1230	CTTCACCAACGTGTACGCCGACAGCTTCGTGATCCGTGGCGACGAGGTGCGGCAGATCGC	1289
Sbjct	1172	CTTTACTAATGTCTATGCAGATTCATTTGTAATTAGAGGTGATGAAGTCAGACAAATCGC	1231
Query	1290	ACCCGGCCAGACAGGCAAGATCGCCGACTACAACCTACAAGCTGCCCGACGACTTCACCGG	1349
Sbjct	1232	TCCAGGGCAAACCTGGAAAGATTGCTGATTATAATTATAAATTACCAGATGATTTTACAGG	1291
Query	1350	CTGCGTGATCGCCTGGAACAGCAACAACCTCGACAGCAAGGTGGGCGGCAACTACAACCTA	1409
Sbjct	1292	CTGCGTTATAGCTTGAATTCTAACAATCTTGATTCTAAGGTTGGTGGTAATTATAATTA	1351
Query	1410	CCTGTACCGGCTGTTCCGGAAGAGCAACCTGAAGCCCTTCGAGCGGGACATCAGCACC GA	1469
Sbjct	1352	CCTGTATAGATTGTTTAGGAAGTCTAATCTCAAACCTTTTGAGAGAGATATTTCAACTGA	1411
Query	1470	GATCTACCAAGCCGGCTCCACCCCTTGCAACGGCGTGGAGGGCTTCAACTGCTACTTCCC	1529
Sbjct	1412	AATCTATCAGGCCGGTAGCACACCTTGTAATGGTGTGAAGGTTTTAATTGTTACTTTCC	1471
Query	1530	TCTGCAGAGCTACGGCTTCCAGCCCACCAACGGCGTGGGCTACCAGCCCTACCGGGTGGT	1589
Sbjct	1472	TTTACAATCATATGGTTTCCAACCCACTAATGGTGTGGTTACCAACCATACAGAGTAGT	1531
Query	1590	GGTGCTGAGCTTCGAGCTGCTGCACGCCCCAGCCACCGTGTGTGGCCCCAAGAAGAGCAC	1649
Sbjct	1532	AGTACTTTCTTTTGAACCTTCTACATGCACCAGCAACTGTTTGTGGACCTAAAAAGTCTAC	1591
Query	1650	CAACCTGGTGAAGAACAAGTGCCTGAACCTTCAACTTCAACGGCCTTACCGGCACCGGCGT	1709
Sbjct	1592	TAATTGTTTAAAAACAAATGTGTCAATTTCAACTTCAATGGTTTAACAGGCACAGGTGT	1651
Query	1710	GCTGACCAGAGCAACAAGAAATTCCTGCCCTTTTCAGCAGTTTCGGCCGGGACATCGCCGA	1769
Sbjct	1652	TCTTACTGAGTCTAACAAAAAGTTTCTGCCTTTCCAACAATTTGGCAGAGACATTGCTGA	1711
Query	1770	CACCACCGACGCTGTGCGGGATCCCCAGACCCTGGAGATCCTGGACATCACCCCTTGCAG	1829
Sbjct	1712	CACTACTGATGCTGTCCGTGATCCACAGACACTTGAGATTCTTGACATTACACCATGTTT	1771
Query	1830	CTTCGGCGGCGTGAGCGTGATCACCCAGGCACCAACACCAGCAACCAGGTGGCCGTGCT	1889
Sbjct	1772	TTTGTTGGTGTGTCAGTGTATATAACACCAGGAACAATACTTCTAACCAGGTTGCTGTTCT	1831
Query	1890	GTACCAGGACGTGAACTGCACCGAGGTGCCCGTGGCCATCCACGCCGACCAGCTGACACC	1949
Sbjct	1832	TTATCAGGATGTAACTGCACAGAAGTCCCTGTTGCTATTTCATGCAGATCAACTTACTCC	1891
Query	1950	CACCTGGCGGGTCTACAGCACCGGCAGCAACGTGTTCCAGACCCGGGCGGGTTGCCTGAT	2009
Sbjct	1892	TACTTGGCGTGTATTATTCTACAGGTTCTAATGTTTTTCAAACACGTGCAGGCTGTTTAAT	1951
Query	2010	CGGCGCCGAGCAGCTGAACAACAGCTACGAGTGCAGATCCCCATCGGCGCCGGCATCTG	2069
Sbjct	1952	AGGGGCTGAACATGTCAACAACCTCATATGAGTGTGACATACCCATTGGTGCAGGTATATG	2011

Query	2070	TGCCAGCTACCAGACCCAGACCAATTACCCCCGAGGGGCAAGGAGCGTGGCCAGCCAGAG	2129
Sbjct	2012	CGCTAGTTATCAGACTCAGACTAATTCTCCTCGGCGGGCACGTAGTGTAGCTAGTCAATC	2071
Query	2130	CATCATCGCCTACACCATGAGCCTGGGCGCCGAGAACAGCGTGGCCTACAGCAACAACAG	2189
Sbjct	2072	CATCATTGCCTACACTATGTCACTTGGTGCAGAAAATTCAGTTGCTTACTCTAATAACTC	2131
Query	2190	CATCGCCATCCCCACCAACTTCACCATCAGCGTGACCACCGAGATTCTGCCCGTGAGCAT	2249
Sbjct	2132	TATTGCCATACCCACAAATTTTACTATTAGTGTTACCACAGAAATTCACAGTGTCTAT	2191
Query	2250	GACCAAGACCAGCGTGGACTGCACCATGTACATCTGCGGGCAGCAGCACCAGTGCAGCAA	2309
Sbjct	2192	GACCAAGACATCAGTAGATTGTACAAATGTACATTGTGGTGATTCAACTGAATGCAGCAA	2251
Query	2310	CCTGCTGTGTCAGTACGGCAGCTTCTGCACCCAGCTGAACCGGGCCCTGACCGGCATCGC	2369
Sbjct	2252	TCTTTTGTGCAATATGGCAGTTTTTGTACACAAATTAACCGTGCTTTAACTGGAATAGC	2311
Query	2370	CGTGAGCAGGACAAGAACACCCAGGAGGTGTTTCGCCAGGTGAAGCAGATCTACAAGAC	2429
Sbjct	2312	TGTTGAACAAGACAAAAACACCAAGAAGTTTTGCACAAGTCAAACAAATTACAAAAAC	2371
Query	2430	CCCTCCCATCAAGGACTTCGGCGGGCTTCAACTTCAGCCAGATCCTGCCCGACCCAGCAA	2489
Sbjct	2372	ACCACCAATTAAAGATTTTGGTGGTTTTAATTTTCACAAATATTACCAGATCCATCAAA	2431
Query	2490	GCCCAGCAAGCGGAGCTTCATCGAGGACCTGCTGTTCAACAAGGTGACCCTAGCCGACGC	2549
Sbjct	2432	ACCAAGCAAGAGGTCTATTATTGAAGATCTACTTTCAACAAAGTGACACTTGCAGATGC	2491
Query	2550	CGGCTTCATCAAGCAGTACGGCGACTGCCTCGGCGACATAGCCGCCCGGGACCTGATCTG	2609
Sbjct	2492	TGGCTTCATCAAAACAATATGGTGATTGCCTTGGTGATATTGCTGCTAGAGACCTCATTG	2551
Query	2610	CGCCCAGAAGTTCAACGGCCTGACCGTGCTGCCTCCCTGCTGACCGACGAGATGATCGC	2669
Sbjct	2552	TGCACAAAAGTTTAAACGGCCTTACTGTTTTGCCACCTTTGCTCACAGATGAAATGATTGC	2611
Query	2670	CCAGTACACCAGCGCCCTGTTAGCCGGAACCATCACCAGCGGCTGGACTTTGGCGCTGG	2729
Sbjct	2612	TCAATACACTTCTGCACTGTTAGCGGGTACAATCACTTCTGGTTGGACCTTTGGTGCAGG	2671
Query	2730	AGCCGCTCTGCAGATCCCTTTCGCCATGCAGATGGCCTACCGGTTCAACGGCATCGGCGT	2789
Sbjct	2672	TGCTGCATTACAAATACCATTGCTATGCAATGGCTTATAGGTTTAAATGGTATTGGAGT	2731
Query	2790	GACCCAGAACGCTGCTGTACGAGAACCAGAAGCTGATCGCCAACCGATTCAACAGCGCCAT	2849
Sbjct	2732	TACACAGAATGTTCTCTATGAGAACCAGAAATGATTGCCAACCAATTTAATAGTGCTAT	2791
Query	2850	CGGCAAGATCCAGGACAGCCTGAGCAGCACCGCTAGCGCCCTGGGCAAGCTGCAGGACGT	2909
Sbjct	2792	TGGCAAAATTCAAGACTCACTTTCTTCCACAGCAAGTGCACTTGGAAAACCTCAAGATGT	2851
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Sbjct	2912	TGCAATTTCAAGTGTTTTAAATGATATCCTTTCACGTCTTGACAAAGTTGAGGCTGAAGT	2971
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Sbjct	2972	GCAAATTGATAGGTTGATCACAGGCAGACTTCAAAGTTTGCAGACATATGTGACTCAACA	3031
Query	3090	GCTGATCCGGGCGCCGAGATTTCGGGCCAGCGCCAACTGGCCGCCACCAAGATGAGCGA	3149
Sbjct	3032	ATTAATTAGAGCTGCAGAAATCAGAGCTTCTGCTAATCTTGCTGCTACTAAAATGTCAGA	3091

Query	3150	GTGCGTIGCTGGGCCAGAGCAAGCGGGTGGACTTCTGCGGCAAGGGCTACACCTGATGAG	3209
Sbjct	3092	GTGTGTACTTGGACAATCAAAAAGAGTTGATTTTTGTGGAAAGGGCTATCATCTTATGTC	3151
Query	3210	CTTTCCCCAGAGCGCACCCCCACGGAGTGGTGTTCCTGCACGTGACCTACGTGCCCGCCCA	3269
Sbjct	3152	CTTCCCTCAGTCAGCACCTCATGGTGTAGTCTTCTGTCATGTGACTTATGTCCCTGCACA	3211
Query	3270	GGAGAAGAACTTCACCACCGCCCCAGCCATCTGCCACGACGGCAAGGCCCACTTTCCCCG	3329
Sbjct	3212	AGAAAAGAACTTCACAACTGCTCCTGCCATTGTGCATGATGGAAAAGCACACTTTCCTCG	3271
Query	3330	GGAGGGCGTGTTCGTGAGCAACGGCACCCACTGGTTCGTGACCCAGCGGAACTTCTACGA	3389
Sbjct	3272	TGAAGGTGTCTTTGTTTCAAATGGCACACACTGGTTTGTAAACAAAGGAATTTTATGA	3331
Query	3390	GCCCCAGATCATCACCACCGACAACACCTTCGTGAGCGGCAACTGCGACGTGGTGATCGG	3449
Sbjct	3332	ACCACAAATCATTACTACAGACAACACATTTGTGTCTGGTAACTGTGATGTTGTAATAGG	3391
Query	3450	CATCGTGAACAACACCGTGTACGATCCCCGTGCAGCCCGAGCTGGACAGCTTCAAGGAGGA	3509
Sbjct	3392	AATTGTCAACAACACAGTTTATGATCCTTTGCAACCTGAATTAGACTCATTCAAGGAGGA	3451
Query	3510	GCTGGACAAGTACTTCAAGAATCACACCAGCCCCGACGTGGACCTGGGCGACATCAGCGG	3569
Sbjct	3452	GTTAGATAAATATTTTAAGAATCATACATCACCAGATGTTGATTTAGGTGACATCTCTGG	3511
Query	3510	GCTGGACAAGTACTTCAAGAATCACACCAGCCCCGACGTGGACCTGGGCGACATCAGCGG	3569
Sbjct	3452	GTTAGATAAATATTTTAAGAATCATACATCACCAGATGTTGATTTAGGTGACATCTCTGG	3511
Query	3570	CATCAACGCCAGCGTGGTGAACATCCAGAAGGAGATCGATCGGCTGAACGAGGTGGCCAA	3629
Sbjct	3512	CATTAATGCTTCAGTTGTAAACATTCAAAAAGAAATTGACCGCCTCAATGAGGTTGCCAA	3571
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Sbjct	3572	GAATTTAAATGAATCTCTCATCGATCTCCAAGAACTTGGAAAAGTATGAGCAGTATATAAA	3631
Query	3690	GTGGCCCTGGTACATCTGGCTGGGCTTCATCGCCGGCCTGATCGCCATCGTGATGGTGAC	3749
Sbjct	3632	ATGGCCATGGTACATTGGCTAGGTTTTATAGCTGGCTTGATTGCCATAGTAATGGTGAC	3691
Query	3750	CATCATgctgtgctgcatgaccagctgctgcagctgcctgaagggctggtgagctgcgg	3809
Sbjct	3692	AATTATGCTTTGCTGTATGACCAGTTGCTGTAGTTGTCTCAAGGGCTGTTGTTCTGTGG	3751
Query	3810	cagctgctgcaAGTTCGACGAGGACGACAGCGAGCCCGTGCTGAAGGGCGTGAAGCTGCA	3869
Sbjct	3752	ATCCTGCTGCAAATTTGATGAAGACGACTCTGAGCCAGTGCTCAAAGGAGTCAAATTACA	3811
Query	3870	CTACAC	3875
Sbjct	3812	TTACAC	3817

Figure 1: Comparison of the mRNA for the S protein in Spikevax (Query) and the mRNA for the S protein in the original Wuhan SARS-CoV-2 (Subject). Vertical lines connect identical bases; the significant difference between the two mRNA sequences is evident.

Query: Figure 2 32321 Spike-encoding contig assembled from Moderna mRNA-1273 vaccine Query ID:

lcl|Query 239423 Length: 4004

>MT192773 Wuhan SARS-CoV-2 reference sequence, S protein

Sequence ID: Query 239425 Length: 3821

Range 1: 36 to 3817

Score:1484 bits (1645), Expect:0.0,

Identities:2603/3786(69%), Gaps:8/3786(0%), Strand: Plus/Plus

Query	56	TGTTTCGTGTTTCTGGTGCTGCTGCCTCTGGTGTCACGCCAGTGTGTGAACCTGACCACCA Sbjct	1	TGTTTTGTTTTTCTTGTTTTATTGCCACTAGTCTCTAGTCAGTGTGTTAATCTTACAACCA	60
Query	116	GAAACACAGCTGCCTCCAGCCTACACCAAACAGCTTTACCAGAGGCGTGTACTACCCCGACA Sbjct	61	GAACTCAATTACCCCCTGCATACTAATTCTTTACACGTGGTGTTTATTACCCTGACA	120
Query	176	AGGTGTTTCAGATCCAGCGTGTCTGCACTCTACCCAGGACCTGTTTCTGCCTTTCTTCAGCA Sbjct	121	AAGTTTTTCAGATCCTCAGTTTTACATTCAACTCAGGACTTGTTCTTACCTTTCTTTTCCA	180
Query	236	ACGTGACCTGGTTCCACGCCATCCACGTGTCCGGCACCAATGGCACCAAGAGATTTCGACA Sbjct	181	ATGTTACTTGGTTCATGCTATACATGTCTCTGGGACCAATGGTACTAAGAGGTTTGATA	240
Query	296	ACCCCGTGCTGCCCTTCAACGACGGGGTGTACTIONTGGCAGCACCGAGAAGTCCAACATCA Sbjct	241	ACCCTGTCTTACCATTTAATGATGGTGTATTATTTGCTTCCACTGAGAACTAACATAA	300
Query	356	TCAGAGGCTGGATCTTTCGGCACCACTGGACAGCAAGACCCAGAGCCTGCTGATCGTGA Sbjct	301	TAAGAGGCTGGATTTTTGGTACTACTTTAGATTCTGAAGACCCAGTCCCTACTTATTGTTA	360
Query	416	ACAACGCCACCAACGTGGTCATCAAAGTGTGCGAGTTCAGTTCGCAACGACCCCTTCC Sbjct	361	ATAACGCTACTAATGTTGTTATTAAAGTCTGTGAATTTCAATTTTGTAATGATCCATTTT	420
Query	476	TGGGCGTCTACTACCACAAGAACAACAAGAGCTGGATGGAAGCGAGTTCCGGGTGTACA Sbjct	421	TGGGTGTTTATTACCACAAAAACA AAAAGTTGGATGGAAGTGAGTTCAGAGTTTATT	480
Query	536	GCAGCGCCAACTGCACCTTCGAGTAGTGTCCCGACCTTTCTGATGGACCTGGAAG Sbjct	481	CTAGTGC GAATAATTGCACTTTTGAATATGTCTCTCAGCCTTTTCTTATGGACCTTGAAG	540
Query	596	GCAAGCAGGGCAACTTCAAGAACCTGCGCGAGTTTCGTGTTTTAAGAACATCGACGGCTACT Sbjct	541	GAAAACAGGGTAATTTCAAAAATCTTAGGGAATTTGTGTTTTAAGAAATATTGATGGTTATT	600
Query	656	TCAAGATCTACAGCAAGCACACCCCTATCAACCTCGTGC GGATCTGCCTCAGGGCTTCT Sbjct	601	TTAAATATATTCTAAGCACACGCCTATTAAATTTAGTGCGTGATCTCCCTCAGGGTTTTT	660
Query	716	CTGCTCTGGAACCCCTGGTGGATCTGCCCATCGGCATCAACATCACCCGTTTCAGACAC Sbjct	661	CGGCTTTAGAACCATTTGGTAGATTG GCAATAGGTATTAACATCACTAGGTTTCAAACCTT	720
Query	776	TGCTGGCCCTGCACAGAAGCTACCTGACACCTGGCGATAGCAGCAGC-GGATGGACAGCT Sbjct	721	TACTTGCTTTACATAGAAGTTATTTGACTCCTGGTGATT-CTTCTTCAGGTTGGACAGCT	779
Query	835	GGTGCCGCGCTTACTATGTGGGCTACCTGCAGCCTAGAACCTTCTGCTGAAGTACAAC Sbjct	780	GGTGCTGCAGCTTATTATGTGGGTTATCTTCAACCTAGGACTTTTCTATTAAATATAAT	839
Query	895	GAGAACGGCACCATCACCGACGCCGTGGATTGTGCTCTGGATCCTCTGAGCGAGACAAAG Sbjct	840	GAAAATGGAACCATACAGATGCTGTAGACTGTGCACTTGACCCTCTCTCAGAAACAAAG	899
Query	955	TGCACCCTGAAGTCCTTACCCTGGAAAAAGGCATCTACCAGACCAGCAACTTCCGGGGTG Sbjct	900	TGTACGTTGAAATCCTTCACTGTAGAAAAAGGAATCTATCAAACCTTCTA ACTTTAGAGTC	959
Query	1015	CAGCCCCACCGAATCCATCGTGCGGTTCCCAATATACCAATCTGTGCCCTTCGGCGAG Sbjct	960	CAACCAACAGAATCTATTGTTAGATTT CCTAATATTACAAACTTG TGCCCTTTTGGTGAA	1019
Query	1075	GTGTTTCAATGCCACCAGATTGCGCTCTGTGTACGCCTGGAACCGGAAGCGGATCAGCAAT Sbjct	1020	GTTTTTAACGCCACCAGATTTCATCTGTTTATGCTTGGAACAGGAAGAGAATCAGCAAC	1077

Query	1135	TGCGTGGCCGACTACTCCGTGCTGTACAACCTCCGC--CAGCTTCAGCACCTTCAAGTGCT	1192
Sbjct	1080	TGTGTTGCTGATTATTCTGTCTATATAATCCGCATCATTTTC--CACTTTTAAGTGTT	1137
Query	1193	ACGGCGTGTCCCCTACCAAGCTGAACGACCTGTGCTTCACAAACGTGTACGCCGACAGCT	1252
Sbjct	1138	ATGGAGTGTCTCCTACTAAATTAAATGATCTCTGCTTTACTAATGTCTATGCAGATTTCAT	1197
Query	1253	TCGTGATCCGGGGAGATGAAGTGCGGCAGATTGCCCTGGACAGACAGGCAAGATCGCCG	1312
Sbjct	1198	TTGTAATTAGAGGTGATGAAGTCAGACAAATCGCTCCAGGGCAAACCTGGAAAGATTGCTG	1257
Query	1313	ACTACAACCTACAAGCTGCCCGACGACTTCACCGGCTGTGTGATTGCCTGGAACAGCAACA	1372
Sbjct	1258	ATTATAATTATAAATTACCAGATGATTTTACAGGCTGCGTTATAGCTTGGAATTCTAACA	1317
Query	1373	ACCTGGACTCCAAAGTCGGCGGCAACTACAATTACCTGTACCGGCTGTTCCGGAAGTCCA	1432
Sbjct	1318	ATCTTGATTCTAAGGTTGGTGGTAATTATAATTACCTGTATAGATTGTTTAGGAAGTCTA	1377
Query	1433	ATCTGAAGCCCTTCGAGCGGGACATCTCCACCGAGATCTATCAGGCCGGCAGCACCCCTT	1492
Sbjct	1378	ATCTCAAACCTTTTGAGAGAGATATTTCAACTGAAATCTATCAGGCCGGTAGCACACCTT	1437
Query	1493	GTAACGGCGTGGAAGGCTTCAACTGCTACTTCCCACTGCAGTCCTACGGCTTTCAGCCCA	1552
Sbjct	1438	GTAATGGTGTGGAAGGTTTTAATTGTTACTTTCTTTACAATCATATGGTTTCCAACCCA	1497
Query	1553	CAAAATGGCGTGGGCTATCAGCCCTACAGAGTGGTGGTGTGAGCTTCGAACTGCTGCATG	1612
Sbjct	1498	CTAATGGTGTGTTGGTTACCAACCATAACAGAGTAGTAGTACTTTCTTTTGAACCTCTACATG	1557
Query	1613	CCCCTGCCACAGTGTGCGGCCCTAAGAAAAGCACCAATCTCGTGAAGAACAAATGCGTGA	1672
Sbjct	1558	CACCAGCAACTGTTTGTGGACCTAAAAAGTCTACTAATTTGGTTAAAAACAAATGTGTCA	1617
Query	1673	ACTTCAACTTCAACGGCCTGACCGGCACCGGCGTGCTGACAGAGAGCAACAAGAAGTTCC	1732
Sbjct	1618	ATTTCAACTTCAATGGTTTAAACAGGCACAGGTGTTCTTACTGAGTCTAACAAAAAGTTTC	1677
Query	1733	TGCCATTCCAGCAGTTTGGCCGGGATATCGCCGATACCACAGACGCCGTTAGAGATCCCC	1792
Sbjct	1678	TGCCTTTCCAACAATTTGGCAGAGACATTGCTGACACTACTGATGCTGTCCGTGATCCAC	1737
Query	1793	AGACACTGGAATCCTGGACATCACCCCTTGACGCTTCGGCGGAGTGTCTGTGATCACCC	1852
Sbjct	1738	AGACACTTGAGATTCTTGACATTACACCATGTTCTTTGGTGGTGTGAGTGTATAACAC	1797
Query	1853	CTGGCACCAACACCAGCAATCAGGTGGCAGTGCTGTACCAGGACGTGAACTGTACCGAAG	1912
Sbjct	1798	CAGGAACAAATACTTCTAACCAGGTTGCTGTTCTTTATCAGGATGTTAACTGCACAGAAG	1857
Query	1913	TGCCCCTGGCCATTACGCGGATCAGCTGACACCTACATGGCGGGTGTACTCCACCGGCA	1972
Sbjct	1858	TCCCTGTTGCTATTTCATGCAGATCAACTTACTCCTACTTGGCGTGTTTATTCTACAGGTT	1917
Query	1973	GCAATGTGTTTTCAGACCAGAGCCGGCTGTCTGATCGGAGCCGAGCACGTGAACAATAGCT	2032
Sbjct	1918	CTAATGTTTTTCAAACACGTGCAGGCTGTTTAAATAGGGGCTGAACATGTCAACAACATCAT	1977
Query	2033	ACGAGTGCAGACATCCCCATCGGCGCTGGAATCTGCGCCAGCTACCAGACACAGACAAACA	2092
Sbjct	1978	ATGAGTGTGACATAACCATTTGGTGCAGGTATATGCGCTAGTTATCAGACTCAGACTAATT	2037
Query	2093	GCCCTCGGAGAGCCAGAAGCGTGGCCAGCCAGAGCATCATTGCCTACACAATGTCTCTGG	2152
Sbjct	2038	CTCCTCGGCGGGCACGTAGTGTAGCTAGTCAATCCATCATTCCTACACTATGTCACTTG	2097

Query	2153	GCGCCGAGAACAGCGTGGCCTACTCCAACAACTCTATCGCTATCCCCACCAACTTCACCA	2212
Sbjct	2098	GTGCAGAAAATTTCAGTTGCTTACTCTAATAACTCTATTGCCATACCCACAAATTTTACTA	2157
Query	2213	TCAGCGTGACCACAGAGATCCTGCCTGTGTCCATGACCAAGACCAGCGTGGACTGCACCA	2272
Sbjct	2158	TTAGTGTTACCACAGAAATTCTACCAGTGTCTATGACCAAGACATCAGTAGATTGTACAA	2217
Query	2273	TGTACATCTGCGGCGATTCCACCGAGTGTCTCCAACCTGCTGCTGCAGTACGGCAGCTTCT	2332
Sbjct	2218	TGTACATTTGTGGTGATTCAACTGAATGCAGCAATCTTTTGTGCAATATGGCAGTTTTT	2277
Query	2333	GCACCCAGCTGAATAGAGCCCTGACAGGGATCGCCGTGGAACAGGACAAGAACACCCAAG	2392
Sbjct	2278	GTACACAATTAAACCGTGCTTTAACTGGAATAGCTGTTGAACAAGACAAAAACACCCAAG	2337
Query	2393	AGGTGTTTCGCCCAAGTGAAGCAGATCTACAAGACCCCTCCTATCAAGGACTTCGGCGGCT	2452
Sbjct	2338	AAGTTTTTGCACAAGTCAAACAAATTTACAAAACACCACCAATTAAAGATTTTGGTGGTT	2397
Query	2453	TCAATTTTCAGCCAGATTCTGCCCCGATCCTAGCAAGCCAGCAAGCGGAGCTTCATCGAGG	2512
Sbjct	2398	TTAATTTTTTCACAAATATTACCAGATCCATCAAAACCAAGCAAGAGGTCATTTATTGAAG	2457
Query	2513	ACCTGCTGTTCAACAAAGTGACACTGGCCGACGCCGGCTTCATCAAGCAGTATGGCGATT	2572
Sbjct	2458	ATCTACTTTTCAACAAAGTGACACTTGACAGATGCTGGCTTCATCAACAATATGGTGATT	2517
Query	2573	GTCTGGGCGACATTGCCGCCAGGGATCTGATTGTGCGCCAGAAAGTTTAACGGACTGACAG	2632
Sbjct	2518	GCCTTGGTGATATTGCTGCTAGAGACCTCATTTGTGCACAAAAGTTTAACGGCCTTACTG	2577
Query	2633	TGCTGCCTCCTCTGCTGACCGATGAGATGATCGCCAGTACACATCTGCCCTGCTGGCCG	2692
Sbjct	2578	TTTTGCCACCTTTGCTCACAGATGAAATGATTGCTCAATACACTTCTGCACTGTTAGCGG	2637
Query	2693	GCACAATCACAAGCGGCTGGACATTTGGAGCAGGCGCCGCTCTGCAGATCCCCTTTGCTA	2752
Sbjct	2638	GTACAATCACCTTCTGGTTGGACCTTTGGTGCAAGGTGCTGCATTACAAATACCATTTGCTA	2697
Query	2753	TGCAGATGGCCTACCGGTTCAACGGCATCGGAGTGACCCAGAATGTGCTGTACGAGAACC	2812
Sbjct	2698	TGCAAAATGGCTTATAGGTTTAATGGTATTGGAGTTACACAGAATGTTCTCTATGAGAACC	2757
Query	2813	AGAAGCTGATCGCCAACCAAGTTCAACAGCGCCATCGGCAAGATCCAGGACAGCCTGAGCA	2872
Sbjct	2758	AAAAATTGATTGCCAACCAATTTAATAGTGCTATTGGCAAAATTCAAGACTCACTTTCTT	2817
Query	2873	GCACAGCAAGCGCCCTGGGAAAGCTGCAGGACGTGGTCAACCAGAATGCCAGGCACTGA	2932
Sbjct	2818	CCACAGCAAGTGCACTTGGAAAACCTTCAAGATGTGGTCAACCAAAATGCACAAGCTTTAA	2877
Query	2933	ACACCCTGGTCAAGCAGCTGTCTCCAACCTTCGGCGCCATCAGCTCTGTGCTGAACGATA	2992
Sbjct	2878	ACACGCTTGTAAACAACCTTAGCTCCAATTTTGGTGCAATTTCAAGTGTTTTAAATGATA	2937
Query	2993	TCCTGAGCAGACTGGACCCTCCTGAGGCGGAGGTGCAGATCGACAGACTGATCACAGGCA	3052
Sbjct	2938	TCCTTTCACGCTTGCACAAAGTTGAGGCTGAAGTGCAAAATTGATAGGTTGATCACAGGCA	2997
Query	3053	GACTGCAGAGCCTCCAGACATACGTGACCCAGCAGCTGATCAGAGCCGCCGAGATTAGAG	3112
Sbjct	2998	GACTTCAAAGTTTGCAGACATATGTGACTCAACAATTAATTAGAGCTGCAGAAATCAGAG	3057
Query	3113	CCTCTGCCAATCTGGCCGCCACCAAGATGTCTGAGTGTGTGCTGGGCCAGAGCAAGAGAG	3172
Sbjct	3058	CTTCTGCTAATCTTGCTGCTACTAAAATGTCAGAGTGTGTACTTGGACAATCAAAAAGAG	3117
Query	3173	TGGACTTTTTCGGCAAGGGCTACCACCTGATGAGCTTCCCTCAGTCTGCCCCCTCACGGCG	3232
Sbjct	3118	TTGATTTTTGTGGAAGGGCTATCATCTTATGTCCTTCCCTCAGTCAGCACCTCATGGTG	3177
Query	3233	TGGTGTCTTGCACGTGACATATGTGCCCGCTCAAGAGAAGAATTTACCACCGCTCCAG	3292
Sbjct	3178	TAGTCTTCTTGCATGTGACTTATGTCCCTGCACAAGAAAAGAACTTCACAACCTGCTCCTG	3237

Query	3293	CCATCTGCCACGACGGCAAAGGCCACTTTTCTAGAGAAGGCCTGTTCGTGTCCAACGGCA 	3352
Sbjct	3238	CCATTGTGTCATGATGGAAAAGCACACTTTCTCGTGAAGGTGTCTTTGTTTTCAAATGGCA	3297
Query	3353	CCCATTGGTTCGTGACACAGCGGAACTTCTACGAGCCCCAGATCATCACCACCGACAACA 	3412
Sbjct	3298	CACACTGGTTTGTAACACAAAGGAATTTTTATGAACCACAAATCATTACTACAGACAACA	335
Query	3413	CCTTCGTGTCTGGCAACTGCGACGTCGTGATCGGCATTGTGAACAATACCGTGTACGACC 	3472
Sbjct	3358	CATTTGTGTCTGGTAACTGTGATGTTGTAATAGGAATTGTCAACAACACAGTTTATGATC	3417
Query	3473	CTCTGCAGCCCGAGCTGGACAGCTTCAAAGAGGAACTGGACAAGTACTTTAAGAACCACA 	3532
Sbjct	3418	CTTTGCAACCTGAATTAGACTCATTCAAGGAGGAGTTAGATAAATATTTAAGAATCATA	3477
Query	3533	CAAGCCCCGACGTGGACCTGGGCGATATCAGCGGAATCAATGCCAGCGTCGTGAACATCC 	3592
Sbjct	3478	CATCACCAGATGTTGATTTAGGTGACATCTCTGGCATTAAATGCTTCAGTTGTAAACATTC	3537
Query	3593	AGAAAGAGATCGACCGGCTGAACGAGGTGGCCAAGAATCTGAACGAGAGCCTGATCGACC 	3652
Sbjct	3538	AAAAAGAAATTGACCGCTCAATGAGGTTGCCAAGAATTTAAATGAATCTCTCATCGATC	3597
Query	3653	TGCAAGAACTGGGGAAGTACGAGCAGTACATCAAGTGGCCCTGGTACATCTGGCTGGGCT 	3712
Sbjct	3598	TCCAAGAACTTGGAAGTATGAGCAGTATATAAATGGCCATGGTACATTGGCTAGGTT	3657
Query	3713	TTATCGCCGACTGATTGCCATCGTGATGGTCACAATCATGCTGTGTTGCATGACCAGCT 	3772
Sbjct	3658	TTATAGCTGGCTTGATTGCCATAGTAATGGTGACAATTATGCTTTGCTGTATGACCAGTT	3717
Query	3773	GCTGTAGCTGCCTGAAGGGCTGTTGTAGCTGTGGCAGCTGCTGCAAGTTCGACGAGGACG 	3832
Sbjct	3718	GCTGTAGTTGTCTCAAGGGCTGTTGTTCTTGTGGATCCTGCTGCAAATTTGATGAAGACG	3777
Query	3833	ATTCTGAGCCCGTGCTGAAGGGCGTGAAACTGCACTACACAT	3874
Sbjct	3778	ACTCTGAGCCAGTGCTCAAAGGAGTCAAATTACATTACACAT	3819

Figure 2: Comparison of S protein mRNA in BNT162b2 (Query) and S protein mRNA in the original Wuhan SARS-CoV-2 (Subject). Vertical lines connect identical bases; a significant difference between the two mRNA sequences is evident.

Query: Figure1_032321_Spike-encoding_contig_assembled_from_BioNTech/Pfizer_BNT-162b2_vaccine; Query ID: lcl|Query_510381 Length: 4175;
>MT192773 Wuhan SARS-CoV-2 reference sequence. S protein

Sequence ID: Query 510383 Length: 3821, Range 1: 1 to 3819

Score:2115 bits (2345), Expect:0.0,

Identities:2765/3822(72%), Gaps:6/3822(0%), Strand: Plus/Plus

Oligonucleotides for quantitative multiplex Real-Time PCR

The mRNA sequences in Spikevax and BNT162b2 differ not only significantly from each other, but also, from the original SARS-CoV-2 mRNA sequence of the original circulating strain “Wuhan” (GenBank MT192773).

These differences were used to distinguish mRNAs from different sources. mRNAs can have different sequences and will still encode

the same protein – the codons (triplets encoding amino acids) can differ for individual amino acids; at least at their third position; codon sequence heterogeneity for amino acids is high.

At the protein level, the S protein in both Spikevax (Moderna) and BNT162b2 (Pfizer) is almost identical and differs from the S protein of the original SARS-CoV-2 “Wuhan” strain that circulated in March 2020 by only two amino acid substitutions, K986P (Lys986Pro) and V987P (Val987Pro). The results are shown in figures 1-5.

Query	50	CCGCCACCATGTTTCGTGTTCTGGTGCTGCTGCCCCCTGGTGAGCAGCCAGTGCCTGAACC	109
Sbjct	47	CCGCCACCATGTTTCGTGTTCTGGTGCTGCTGCCCCCTGGTGAGCAGCCAGTGCCTGAACC	106
Query	110	TGACCACCCGGACCCAGCTGCCACCAGCCTACACCAACAGCTTCACCCGGGGCGTCTACT	169
Sbjct	107	TGACCACCAGAACACAGCTGCCTCCAGCCTACACCAACAGCTTTACCAGAGGCGTGTACT	166
Query	170	ACCCCGACAAGGTGTTCCGGAGCAGCGTCTGCACAGCACCAGGACCTGTTCTGCCCT	229
Sbjct	167	ACCCCGACAAGGTGTTCCAGATCCAGCGTGTGCACTCTACCCAGGACCTGTTCTGCCCT	226
Query	230	TCTTCAGCAACGTGACCTGGTTCACGCCATCCACGTGAGCGGCACCAACGGCACCAGC	289
Sbjct	227	TCTTCAGCAACGTGACCTGGTTCACGCCATCCACGTGTCCGGCACCAGTGGCACCAGC	286
Query	290	GGTTCGACAACCCCGTGTGCTGCCCTTCAACGACGGCGTGTACTTCGCCAGCACCAGAGA	349
Sbjct	287	GATTCGACAACCCCGTGTGCTGCCCTTCAACGACGGGGTGTACTTTGCCAGCACCAGAGA	346
Query	350	GCAACATCATCCGGGGCTGGATCTTCGGCACCACCCTGGACAGCAAGACCCAGAGCCTGC	409
Sbjct	347	CCAACATCATCAGAGGCTGGATCTTCGGCACCACACTGGACAGCAAGACCCAGAGCCTGC	406
Query	410	TGATCGTGAATAACGCCACCAACGTGGTGATCAAGGTGTGCGAGTTCCAGTTCTGCAACG	469
Sbjct	407	TGATCGTGAATAACGCCACCAACGTGGTGATCAAGGTGTGCGAGTTCCAGTTCTGCAACG	466
Query	470	ACCCCTTCCTGGGCGTGTACTACCACAAGAACAACAAGAGCTGGATGGAGAGCGAGTTCC	529
Sbjct	467	ACCCCTTCCTGGGCGTGTACTACCACAAGAACAACAAGAGCTGGATGGAAAGCGAGTTCC	526
Query	530	GGGTGTACAGCAGCGCCAACAACCTGCACCTTCGAGTACGTGAGCCAGCCCTTCCTGATGG	589
Sbjct	527	GGGTGTACAGCAGCGCCAACAACCTGCACCTTCGAGTACGTGTCCAGCCTTCCTGATGG	586
Query	590	ACCTGGAGGGCAAGCAGGGCAACTTCAAGAACCTGCGGGAGTTCTGTTCAGAATCG	649
Sbjct	587	ACCTGGAAGGCAAGCAGGGCAACTTCAAGAACCTGCGCGAGTTCTGTTCAGAATCG	646
Query	650	ACGGCTACTTCAAGATCTACAGCAAGCACACCCCAATCAACCTGGTGCGGGATCTGCCCT	709
Sbjct	647	ACGGCTACTTCAAGATCTACAGCAAGCACACCCCTATCAACCTCGTGCGGGATCTGCCCT	706
Query	710	AGGGCTTCTCAGCCCTGGAGCCCTGGTGGACCTGCCCATCGGCATCAACATCACCCGGT	769
Sbjct	707	AGGGCTTCTCAGCCCTGGAGCCCTGGTGGACCTGCCCATCGGCATCAACATCACCCGGT	766
Query	770	TCCAGACCCTGCTGGCCCTGCACCGGAGCTACCTGACCCCAGGCGACAGCAGCAGCGGGT	829
Sbjct	767	TTCAGACACTGCTGGCCCTGCACAGAAGCTACCTGACACCTGGCGATAGCAGCAGCGGAT	826
Query	830	GGACAGCAGGCGCGGCTGCTTACTACGTGGGCTACCTGCAGCCCCGGACCTTCCTGCTGA	889
Sbjct	827	GGACAGCTGGTGCCCGCGCTTACTATGTGGGCTACCTGCAGCCTAGAACCTTCCTGCTGA	886
Query	890	AGTACAACGAGAACGGCACCATCACCGACGCCGTGGACTGCGCCCTGGACCTCTGAGCG	949
Sbjct	887	AGTACAACGAGAACGGCACCATCACCGACGCCGTGGATTGTGCTCTGGATCCTCTGAGCG	946

Query	950	AGACCAAGTGCACCCTGAAGAGCTTCACCGTGGAGAAGGGCATCTACCAGACCAGCAACT	1009
Sbjct	947	AGACAAAGTGCACCCTGAAGTCCTTCACCGTGGAAAAGGGCATCTACCAGACCAGCAACT	1006
Query	1010	TCCGGGTGCAGCCCACCGAGAGCATCGTGCGGTTCCTCAACATCACCAACCTGTGCCCCCT	1069
Sbjct	1007	TCCGGGTGCAGCCCACCGAATCCATCGTGCGGTTCCTCAATATCACCAATCTGTGCCCCCT	1066
Query	1070	TCGGCGAGGTGTTCAACGCCACCCGGTTCGCCAGCGTGTACGCCTGGAACCGGAAGCGGA	1129
Sbjct	1067	TCGGCGAGGTGTTCAATGCCACCAGATTGCCTCTGTGTACGCCTGGAACCGGAAGCGGA	1126
Query	1130	TCAGCAACTGCGTGGCCGACTACAGCGTGTGTACAACAGCGCCAGCTTCAGCACCTTCA	1189
Sbjct	1127	TCAGCAATTGCGTGGCCGACTACTCCGTGTGTACAACCTCCGCCAGCTTCAGCACCTTCA	1186
Query	1190	AGTGCTACGGCGTGAGCCCCACCAAGCTGAACGACCTGTGCTTCACCAACGTGTACGCCG	1249
Sbjct	1187	AGTGCTACGGCGTGTCCCTACCAAGCTGAACGACCTGTGCTTCACAAACGTGTACGCCG	1246
Query	1250	ACAGCTTCGTGATCCGTGGCGACGAGGTGCGGCAGATCGCACCCGGCCAGACAGGCAAGA	1309
Sbjct	1247	ACAGCTTCGTGATCCGGGGAGATGAAGTGCGGCAGATTGCCCTGGACAGACAGGCAAGA	1306
Query	1310	TCGCCGACTACAACCTACAAGCTGCCCGACGACTTCACCGGCTGCGTGATCGCCTGGAACA	1369
Sbjct	1307	TCGCCGACTACAACCTACAAGCTGCCCGACGACTTCACCGGCTGTGTGATTGCCTGGAACA	1366
Query	1370	GCAACAACCTCGACAGCAAGGTGGGCGGCAACTACAACCTACCTGTACCGGCTGTTCCGGA	1429
Sbjct	1367	GCAACAACCTGGACTCCAAAGTCGGCGGCAACTACAATTACCTGTACCGGCTGTTCCGGA	1426
Query	1430	AGAGCAACCTGAAGCCCTTCGAGCGGGACATCAGCACCGAGATCTACCAAGCCGGCTCCA	1489
Sbjct	1427	AGTCCAATCTGAAGCCCTTCGAGCGGGACATCTCCACCGAGATCTATCAGGCCGGCAGCA	1486
Query	1490	CCCTTGCAACGGCGTGGAGGGCTTCAACTGCTACTTCCCTCTGCAGAGCTACGGCTTCC	1549
Sbjct	1487	CCCTTGTAACGGCGTGGAAAGGCTTCAACTGCTACTTCCCACTGCAGTCTACGGCTTTC	1546
Query	1550	AGCCCAACCAACGGCGTGGGCTACCAGCCCTACCGGGTGGTGGTGCTGAGCTTCGAGCTGC	1609
Sbjct	1547	AGCCCACAAATGGCGTGGGCTATCAGCCCTACAGAGTGGTGGTGCTGAGCTTCGAACTGC	1606
Query	1610	TGCACGCCCCAGCCACCGTGTGTGGCCCCAAGAAGAGCACCAACCTGGTGAAGAACAAGT	1669
Sbjct	1607	TGCATGCCCTGCCACAGTGTGCGGCCCTAAGAAAAGCACCAATCTCGTGAAGAACAAT	1666
Query	1670	GCGTGAACCTCAACTTCAACGGCCTTACCGGCACCGGCGTGTGACCGAGAGCAACAAGA	1729
Sbjct	1667	GCGTGAACCTCAACTTCAACGGCCTGACCGGCACCGGCGTGTGACAGAGAGCAACAAGA	1726
Query	1730	AATTCCTGCCCTTTTACGAGTTTCGGCCGGGACATCGCCGACACACCGACGCTGTGCGGG	1789
Sbjct	1727	AGTTCTGCCATTCCAGCAGTTTGGCCGGGATATCGCCGATACCACAGACGCCGTTAGAG	1786
Query	1790	ATCCCCAGACCCTGGAGATCCTGGACATCACCCCTTGCAGCTTCGGCGGCGTGAGCGTGA	1849
Sbjct	1787	ATCCCCAGACACTGGAATCCTGGACATCACCCCTTGCAGCTTCGGCGGAGTGTCTGTGA	1846

Query	1850	TCACCCAGGCACCAACACCAGCAACCAGGTGGCCGTGCTGTACCAGGACGTGAACTGCA	1909
Sbjct	1847	TCACCCCTGGCACCAACACCAGCAATCAGGTGGCAGTGCTGTACCAGGACGTGAACTGTA	1906
Query	1910	CCGAGGTGCCCCTGGCCATCCACGCCGACCAGCTGACACCCACCTGGCGGGTCTACAGCA	1969
Sbjct	1907	CCGAAGTGCCCCTGGCCATTACGCCGATCAGCTGACACCTACATGGCGGGTGTACTCCA	1966
Query	1970	CCGGCAGCAACGTGTTCCAGACCCGGGCGGTTGCCTGATCGGCGCCGAGCACGTGAACA	2029
Sbjct	1967	CCGGCAGCAATGTGTTTCAGACCAGAGCCGGCTGTCTGATCGGAGCCGAGCACGTGAACA	2026
Query	2030	ACAGCTACGAGTGCGACATCCCCATCGGCGCCGGCATCTGTGCCAGCTACCAGACCCAGA	2089
Sbjct	2027	ATAGCTACGAGTGCGACATCCCCATCGGCGCTGGAATCTGCGCCAGCTACCAGACACAGA	2086
Query	2090	CCAATTCACCCCGGAGGGCAAGGAGCGTGCCAGCCAGAGCATCATCGCTACACCATGA	2149
Sbjct	2087	CAAACAGCCCTCGGAGAGCCAGAAGCGTGCCAGCCAGAGCATCATTGCCTACACAATGT	2146
Query	2150	GCCTGGGCGCCGAGAACAGCGTGGCCTACAGCAACAACAGCATCGCCATCCCCACCAACT	2209
Sbjct	2147	CTCTGGGCGCCGAGAACAGCGTGGCCTACTCCAACAACTCTATCGCTATCCCCACCAACT	2206
Query	2210	TCACCATCAGCGTGACCACCGAGATTCTGCCCCTGAGCATGACCAAGACCAGCGTGGACT	2269
Sbjct	2207	TCACCATCAGCGTGACCACAGAGATCCTGCCTGTGTCCATGACCAAGACCAGCGTGGACT	2266
Query	2270	GCACCATGTACATCTGCGGCGACAGCACCAGTGTCAGCAACCTGCTGCTGCAGTACGGCA	2329
Sbjct	2267	GCACCATGTACATCTGCGGCGATTCCACCAGTGCTCCAACCTGCTGCTGCAGTACGGCA	2326
Query	2330	GCTTCTGCACCCAGCTGAACCGGGCCCTGACCGGCATCGCCGTGGAGCAGGACAAGAACA	2389
Sbjct	2327	GCTTCTGCACCCAGCTGAATAGAGCCCTGACAGGGATCGCCGTGGAACAGGACAAGAACA	2386
Query	2390	CCCAGGAGGTGTTTCGCCAGGTGAAGCAGATCTACAAGACCCCTCCCATCAAGGACTTCG	2449
Sbjct	2387	CCCAAGAGGTGTTTCGCCAAGTGAAGCAGATCTACAAGACCCCTCCTATCAAGGACTTCG	2446
Query	2450	GCGGCTTCAACTTCAGCCAGATCCTGCCCCAGCCAGCAAGCCCAGCAAGCGGAGCTTCA	2509
Sbjct	2447	GCGGCTTCAATTTTCAGCCAGATTCTGCCCCATCTAGCAAGCCCAGCAAGCGGAGCTTCA	2506
Query	2510	TCGAGGACCTGCTGTTCAACAAGGTGACCCTAGCCGACGCCGGCTTCATCAAGCAGTACG	2569
Sbjct	2507	TCGAGGACCTGCTGTTCAACAAGGTGACACTGGCCGACGCCGGCTTCATCAAGCAGTATG	2566
Query	2570	GCGACTGCCTCGGCGACATAGCCGCCCGGGACCTGATCTGCGCCAGAAAGTTCAACGGCC	2629
Sbjct	2567	GCGATTGTCTGGGCGACATTGCCGCCAGGGATCTGATTGCGCCAGAAAGTTTAACGGAC	2626
Query	2630	TGACCGTGCTGCCTCCCCTGCTGACCGACGAGATGATCGCCAGTACACCAGCGCCCTGT	2689
Sbjct	2627	TGACAGTGCTGCCTCCTCTGCTGACCGATGAGATGATCGCCAGTACACATCTGCCCTGC	2686
Query	2690	TAGCCGGAACCATCACCAGCGGCTGGACTTTCGGCGCTGGAGCCGCTCTGCAGATCCCCT	2749
Sbjct	2687	TGGCCGGCACAATCACAAGCGGCTGGACATTTGGAGCAGGCGCCGCTCTGCAGATCCCCT	2746
Query	2750	TCGCCATGCAGATGGCCTACCGGTTCAACGGCATCGGCGTGACCCAGAAACGTGCTGTACG	2809
Sbjct	2747	TTGCTATGCAGATGGCCTACCGGTTCAACGGCATCGGAGTGACCCAGAAATGTGCTGTACG	2806

Query	2810	AGAACCAGAAGCTGATCGCCAACCAGTTCAACAGCGCCATCGGCAAGATCCAGGACAGCC	2869
Sbjct	2807	AGAACCAGAAGCTGATCGCCAACCAGTTCAACAGCGCCATCGGCAAGATCCAGGACAGCC	2866
Query	2870	TGAGCAGCACCGCTAGCGCCCTGGGCAAGCTGCAGGACGTGGTGAACCAGAACGCCCCAGG	2929
Sbjct	2867	TGAGCAGCACAGCAAGCGCCCTGGGAAAGCTGCAGGACGTGGTCAACCAGAATGCCCCAGG	2926
Query	2930	CCCTGAACACCCTGGTGAAGCAGCTGAGCAGCAACTTCGGCGCCATCAGCAGCGTGCTGA	2989
Sbjct	2927	CACTGAACACCCTGGTCAAGCAGCTGTCTTCCAACCTTCGGCGCCATCAGCTCTGTGCTGA	2986
Query	2990	ACGACATCCTGAGCCGGCTGGACCCTCCCGAGGCCGAGGTGCAGATCGACCGGCTGATCA	3049
Sbjct	2987	ACGATATCCTGAGCAGACTGGACCCTCCTGAGGCCGAGGTGCAGATCGACAGACTGATCA	3046
Query	3050	CTGGCCGGCTGCAGAGCCTGCAGACCTACGTGACCCAGCAGCTGATCCGGGCCGCCGAGA	3109
Sbjct	3047	CAGGCAGACTGCAGAGCCTCCAGACATACGTGACCCAGCAGCTGATCAGAGCCGCCGAGA	3106
Query	3110	TTCGGGCCAGCGCCAACCTGGCCGCCACCAAGATGAGCGAGTGCGTGCTGGGCCAGAGCA	3169
Sbjct	3107	TTAGAGCCTCTGCCAATCTGGCCGCCACCAAGATGTCTGAGTGTGTGCTGGGCCAGAGCA	3166
Query	3170	AGCGGGTGGACTTCTGCGGCAAGGGCTACCACCTGATGAGCTTTCCCCAGAGCGCACCCC	3229
Sbjct	3167	AGAGAGTGGACTTTTGGCGCAAGGGCTACCACCTGATGAGCTTCCCTCAGTCTGCCCTC	3226
Query	3230	ACGGAGTGGTGTTCCTGCACGTGACCTACGTGCCCCGCCAGGAGAAGAACTTCACCACCG	3289
Sbjct	3227	ACGGCGTGGTGTTCCTGCACGTGACATATGTGCCCGCTCAAGAGAAGAAATTCACCACCG	3286
Query	3290	CCCCAGCCATCTGCCACGACGGCAAGGCCCACTTTCCCGGGAGGGCGTGTTCGTGAGCA	3349
Sbjct	3287	CTCCAGCCATCTGCCACGACGGCAAGGCCCACTTTCCCTAGAGAAGGCGTGTTCGTGTCCA	3346
Query	3350	ACGGCACCCACTGGTTTCGTGACCCAGCGGAACTTCTACGAGCCCCAGATCATCACCACCG	3409
Sbjct	3347	ACGGCACCCATTGGTTTCGTGACACAGCGGAACTTCTACGAGCCCCAGATCATCACCACCG	3406
Query	3410	ACAACACCTTCGTGAGCGGCAACTGCGACGTGGTGATCGGCATCGTGAACAACACCGTGT	3469
Sbjct	3407	ACAACACCTTCGTGTCTGGCAACTGCGACGTGCTGATCGGCATTGTGAACAATACCGTGT	3466
Query	3470	ACGATCCCCTGCAGCCCAGCTGGACAGCTTCAAGGAGGAGCTGGACAAGTACTTCAAGA	3529
Sbjct	3467	ACGACCCTCTGCAGCCCAGCTGGACAGCTTCAAGAGGAACTGGACAAGTACTTTAAGA	3526
Query	3530	ATCACACCAGCCCCGACGTGGACCTGGGCGACATCAGCGGCATCAACGCCAGCGTGGTGA	3589
Sbjct	3527	ACCACACAAGCCCCGACGTGGACCTGGGCGATATCAGCGGAATCAATGCCAGCGTCTGTA	3586
Query	3590	ACATCCAGAAGGAGATCGATCGGCTGAACGAGGTGGCCAAGAACCTGAACGAGAGCCTGA	3649
Sbjct	3587	ACATCCAGAAAGAGATCGACCGGCTGAACGAGGTGGCCAAGAATCTGAACGAGAGCCTGA	3646
Query	3650	TCGACCTGCAGGAGCTGGGCAAGTACGAGCAGTACATCAAGTGGCCCTGGTACATCTGGC	3709
Sbjct	3647	TCGACCTGCAAGAACTGGGGAAGTACGAGCAGTACATCAAGTGGCCCTGGTACATCTGGC	3706
Query	3710	TGGGCTTCATCGCCGGCCTGATCGCCATCGTGATGGTGACCATCAgtctgtgctgcatga	3769
Sbjct	3707	TGGGCTTTATCGCCGGACTGATTGCCATCGTGATGGTCACAATCATGCTGTGTGTCATGA	3766


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Query    3770    ccagctgctgcagctgcctgaagggtgttgcagctgcggcagctgctgcaAGTTCGACG    3829
          |||
Sbjct    3767    CCAGCTGCTGTAGCTGCCTGAAGGGCTGTTGTAGCTGTGGCAGCTGCTGCAAGTTCGACG    3826

Query    3830    AGGACGACAGCGAGCCCGTGCTGAAGGGCGTGAAGCTGCACTACACCTGAT    3880
          |||
Sbjct    3827    AGGACGATTCTGAGCCCGTGCTGAAGGGCGTGAAGCTGCACTACACATGAT    3877

```

Figure 3: Comparison of mRNA for S protein in Spikevax (Query) and mRNA for S protein in BNT162b2 (Subject). Vertical lines connect identical bases; a significant difference between the two mRNA sequences is evident.

Query: Figure 2 32321 Spike-encoding contig assembled from Moderna mRNA-1273 vaccine Query ID: lcl|Query 246871 Length: 4004

>Figure1 032321 Spike-encoding contig assembled from BioNTech/Pfizer BNT-162b2 vaccine

Sequence ID: Query 246873 Length: 4175, Range 1: 47 to 3877

Score:5273 bits (5847), Expect:0.0,

Identities:3468/3831(91%), Gaps:0/3831(0%), Strand: Plus/Plus

Query	1	MFVFLVLLPLVSSQCVNLTTTRTQLPPAYTNSFTRGVYYPDKVFRSSVLHSTQDLFLPFFS	60
Sbjct	1	MFVFLVLLPLVSSQCVNLTTTRTQLPPAYTNSFTRGVYYPDKVFRSSVLHSTQDLFLPFFS	60
Query	61	NVTWFHAIHVSNGTKRFDNPVLPFNDGVYFASTSEKSNIRGWIFGTTLDSKTQSLIIV	120
Sbjct	61	NVTWFHAIHVSNGTKRFDNPVLPFNDGVYFASTSEKSNIRGWIFGTTLDSKTQSLIIV	120
Query	121	NNATNVVIKVFCEQFCNDPFLGVYHKNKSWMESEFRVYSSANNCTFEYVSQPFMDLE	180
Sbjct	121	NNATNVVIKVFCEQFCNDPFLGVYHKNKSWMESEFRVYSSANNCTFEYVSQPFMDLE	180
Query	181	GKQGNFKNLREFVFNIDGYFKIYSKHTPINLVRDLPQGFSALEPLVDLPIGINITRFQT	240
Sbjct	181	GKQGNFKNLREFVFNIDGYFKIYSKHTPINLVRDLPQGFSALEPLVDLPIGINITRFQT	240
Query	241	LLALHRSYLTTPGDSSSGWTAGAAAYYVGYLQPRFTLLKYNENGTITDAVDCALDPLSETK	300
Sbjct	241	LLALHRSYLTTPGDSSSGWTAGAAAYYVGYLQPRFTLLKYNENGTITDAVDCALDPLSETK	300
Query	301	CTLKSFTVEKGIYQTSNFRVQPTESIVRFPNITNLCPFGEVFNATRFASVYAWNRRKRISN	360
Sbjct	301	CTLKSFTVEKGIYQTSNFRVQPTESIVRFPNITNLCPFGEVFNATRFASVYAWNRRKRISN	360
Query	361	CVADYSVLYNSASFSTFKCYGVSPTKLNDLCFTNVYADSFVIRGDEVRQIAPGQTGKIAD	420
Sbjct	361	CVADYSVLYNSASFSTFKCYGVSPTKLNDLCFTNVYADSFVIRGDEVRQIAPGQTGKIAD	420
Query	421	YNYKLPDDFTGCVIAWNSNNLDSKVGNNYLYRLFRKSNLKPFERDISTEYQAGSTPC	480
Sbjct	421	YNYKLPDDFTGCVIAWNSNNLDSKVGNNYLYRLFRKSNLKPFERDISTEYQAGSTPC	480
Query	481	NGVEGFNCYFPLQSYGFQPTNGVGYQPYRVVLSFELLHAPATVCGPKKSTNLVKNKCVN	540
Sbjct	481	NGVEGFNCYFPLQSYGFQPTNGVGYQPYRVVLSFELLHAPATVCGPKKSTNLVKNKCVN	540
Query	541	FNFNGLTGTGVLTESNKKFLPFQFGRDIADTTDAVRDPQTLEILDITPCSFGGVSVITP	600
Sbjct	541	FNFNGLTGTGVLTESNKKFLPFQFGRDIADTTDAVRDPQTLEILDITPCSFGGVSVITP	600

Query	601	GTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGSNVFQTRAGCLIGAEHVNNNSY	660
Sbjct	601	GTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGSNVFQTRAGCLIGAEHVNNNSY	660
Query	661	ECDIPIGAGICASYQTQTNSPRRARSVASQSIIAYTMSLGAENSVAYSNNNSIAIPTNFTI	720
Sbjct	661	ECDIPIGAGICASYQTQTNSPRRARSVASQSIIAYTMSLGAENSVAYSNNNSIAIPTNFTI	720
Query	721	SVTTEILPVSMTKTSVDCTMYICGDSTECSNLLQYGSFCTQLNRALTGIAVEQDKNTQE	780
Sbjct	721	SVTTEILPVSMTKTSVDCTMYICGDSTECSNLLQYGSFCTQLNRALTGIAVEQDKNTQE	780
Query	781	VFAQVKQIYKTPPIKDFGGFNFSQILPDPSKPSKRSFIEDLLFNKVTLDAGFIKQYGDC	840
Sbjct	781	VFAQVKQIYKTPPIKDFGGFNFSQILPDPSKPSKRSFIEDLLFNKVTLDAGFIKQYGDC	840
Query	841	LGDI AARDLICAQKFNGLT VLPPLLTDEMIAQYTSALLAGTITSGWTFGAGAALQIPFAM	900
Sbjct	841	LGDI AARDLICAQKFNGLT VLPPLLTDEMIAQYTSALLAGTITSGWTFGAGAALQIPFAM	900
Query	901	QMA YRFNGIGVTQNVLYENQKLIANQFN SAIGKIQDSLSTASALGKLQDVVNQNAQALN	960
Sbjct	901	QMA YRFNGIGVTQNVLYENQKLIANQFN SAIGKIQDSLSTASALGKLQDVVNQNAQALN	960
Query	961	TLVKQLSSNFGAISSVLNDILSRLDPEAEVQIDRLITGRLQSLQTYVTQQLIRAAEIRA	1020
Sbjct	961	TLVKQLSSNFGAISSVLNDILSRLDPEAEVQIDRLITGRLQSLQTYVTQQLIRAAEIRA	1020
Query	1021	SANLAATKMSECVLGQSKRVDFCGKGYHLMSFPQSAPHGVVFLHVTYVPAQEKNTTAPA	1080
Sbjct	1021	SANLAATKMSECVLGQSKRVDFCGKGYHLMSFPQSAPHGVVFLHVTYVPAQEKNTTAPA	1080
Query	1081	ICHDKGAHFPREGVFVSNGTWHFVTQRNFYEPQIIITDNTFVSGNCDVVIGIVNNTVYDP	1140
Sbjct	1081	ICHDKGAHFPREGVFVSNGTWHFVTQRNFYEPQIIITDNTFVSGNCDVVIGIVNNTVYDP	1140
Query	1141	LQPELDSFKEELDKYFKNHTSPDVLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDL	1200
Sbjct	1141	LQPELDSFKEELDKYFKNHTSPDVLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDL	1200
Query	1201	QELGKYEQYIKWPWYIWLGFIAGLIAIVMTIMLCCMTSCCSCLKGCCSCGSCCKFDEDD	1260
Sbjct	1201	QELGKYEQYIKWPWYIWLGFIAGLIAIVMTIMLCCMTSCCSCLKGCCSCGSCCKFDEDD	1260
Query	1261	SEPVLKGVKLHYT	1273
Sbjct	1261	SEPVLKGVKLHYT	1273

Figure 4: Comparison of the translated S protein in Spikevax (Query) and the translated S protein in the original Wuhan SARS-CoV-2 (Subject). With the exception of two amino acid substitutions in Spikevax (K986P; Lys986Pro and V987P; Val987Pro), which are highlighted in red, the two sequences are identical at the protein level.

Query: Moderna S protein ID: lcl|Query_22211 Length: 1273

>Sbjct: Wuhan S protein

Sequence ID: Query_22213 Length: 1273

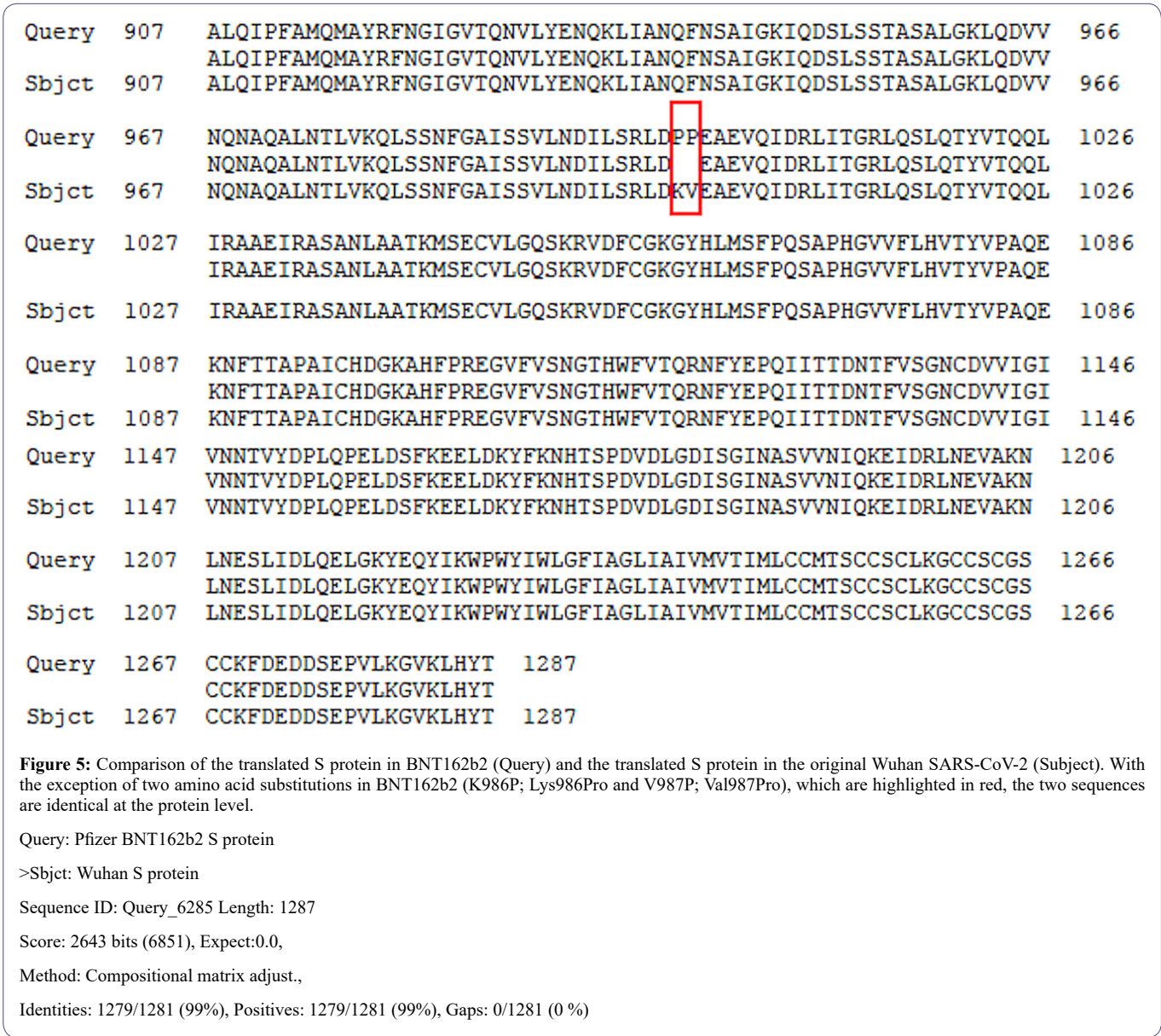
Range 1: 1 to 1273

Score:2630 bits (6818), Expect:0.0,

Method: Compositional matrix adjust.

Identities:1271/1273(99%), Positives:1271/1273(99%), Gaps:0/1273(0%)

Query	1	MFVFLVLLPLVSSQCVNLTTTRTQLPPAYTNSFTRGVYYPDKVFRSSVLHSTQ	66
Sbjct	1	MFVFLVLLPLVSSQCVNLTTTRTQLPPAYTNSFTRGVYYPDKVFRSSVLHSTQ	66
Query	67	DLFLPFFSNVTWFHAIHVSGTNGTKRFDNPVLPFNDGVYFASTEKSNIIRGWIFGTTLDS	126
Sbjct	67	DLFLPFFSNVTWFHAIHVSGTNGTKRFDNPVLPFNDGVYFASTEKSNIIRGWIFGTTLDS	126
Query	127	KTQSLIVNNATNVVIKVFCEQFCNDPFLGVYYHKNNKSWMESEFRVYSSANNCTFEYVS	186
Sbjct	127	KTQSLIVNNATNVVIKVFCEQFCNDPFLGVYYHKNNKSWMESEFRVYSSANNCTFEYVS	186
Query	187	QPFLMDLEGKQGNFKNLREFVFKNIDGYFKIYSKHTPINLVRDLPGQFSALEPLVDLP	246
Sbjct	187	QPFLMDLEGKQGNFKNLREFVFKNIDGYFKIYSKHTPINLVRDLPGQFSALEPLVDLP	246
Query	247	INITRFQTLALHRSYLTGPDSSSGWTAGAAAYVGYLQPRTFLLKYNENGTITDAVDCA	306
Sbjct	247	INITRFQTLALHRSYLTGPDSSSGWTAGAAAYVGYLQPRTFLLKYNENGTITDAVDCA	306
Query	307	LDPLSETKCTLKSFTVEKGIYQTSNFRVQPTESIVRFPNITNLCPFGEVFNATRFASVYA	366
Sbjct	307	LDPLSETKCTLKSFTVEKGIYQTSNFRVQPTESIVRFPNITNLCPFGEVFNATRFASVYA	366
Query	367	WNRKRISNCVADYSVLYNSASFSTFKCYGVSPTKLNDLCFTNVYADSFVIRGDEVROIAP	426
Sbjct	367	WNRKRISNCVADYSVLYNSASFSTFKCYGVSPTKLNDLCFTNVYADSFVIRGDEVROIAP	426
Query	427	GQTGKIADYNYKLPDDFTGCVIAWNSNNLDSKVGNNYNYLYRLFRKSNLKPFERDISTEI	486
Sbjct	427	GQTGKIADYNYKLPDDFTGCVIAWNSNNLDSKVGNNYNYLYRLFRKSNLKPFERDISTEI	486
Query	487	YQAGSTPCNGVEGFNCYFPLQSYGFQPTNGVGYQPYRVVVLSEFLLHAPATVCGPKKSTN	546
Sbjct	487	YQAGSTPCNGVEGFNCYFPLQSYGFQPTNGVGYQPYRVVVLSEFLLHAPATVCGPKKSTN	546
Query	547	LVKNKCVNFNFNGLTGTGVLTESNKKFLPFQFQGRDIADTTDAVRDPQTLIELDITPCSF	606
Sbjct	547	LVKNKCVNFNFNGLTGTGVLTESNKKFLPFQFQGRDIADTTDAVRDPQTLIELDITPCSF	606
Query	607	GGVSVITPGTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGNSNVFQTRAGCLIG	666
Sbjct	607	GGVSVITPGTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGNSNVFQTRAGCLIG	666
Query	667	AEHVNNSEYCDIPIGAGICASYQTQTNPRRARSVASQSIIAYTMSLGAENSVAYSNNNSI	726
Sbjct	667	AEHVNNSEYCDIPIGAGICASYQTQTNPRRARSVASQSIIAYTMSLGAENSVAYSNNNSI	726
Query	727	AIPTNFTISVTTEILPVSMTKTSVDCTMYICGDSTECSNLLQYGSFCTQLNRALTGIAV	786
Sbjct	727	AIPTNFTISVTTEILPVSMTKTSVDCTMYICGDSTECSNLLQYGSFCTQLNRALTGIAV	786
Query	787	EQDKNTQEVFAQVKQIYKTPPIKDFGGFNFSQILPDPSKPSKRSFIEDLLFNKVTILADAG	846
Sbjct	787	EQDKNTQEVFAQVKQIYKTPPIKDFGGFNFSQILPDPSKPSKRSFIEDLLFNKVTILADAG	846
Query	847	FIKQYGDCLGDIAARDLICAQKFNGLTIVLPPLLTDEMIAQYTSALLAGTITSGWTFGAGA	906
Sbjct	847	FIKQYGDCLGDIAARDLICAQKFNGLTIVLPPLLTDEMIAQYTSALLAGTITSGWTFGAGA	906



Based upon the bioinformatic analysis a region was identified in the mRNA sequence of Spikevax and BNT162b2 that was sufficiently homologous to allow primers to be designed for amplification of both Spikevax and BNT162b2 S protein mRNAs, and at the same time sufficiently heterologous to allow fluorescently labeled hybridization probes to accurately discriminate between Spikevax and BNT162b2 S protein mRNAs by quantitative Real-Time PCR. The region that was selected is shown in figure 6.

Primers and fluorescently labelled probe for quantitative Real-Time PCR detection of *Escherichia coli* genomic DNA (HEX) were designed to the internal transcribed spacer region (ITS) of the 16S rDNA cassette. The assay has been validated according to ISO 13485 and is routinely diagnostically used by the laboratory.

All primers and fluorescently labeled hybridization probes used were custom synthesized by Eurofins Genomics, DE.

The sequences of the primers and fluorescently labeled hybridization probes used in the multiplex quantitative Real-Time PCR of mRNA and expression cassettes for S protein Spikevax, BNT162b2, expression cloning vector and *Escherichia coli* genomic DNA are shown in table 1.

Primers and the fluorescently labeled probe (Cy5) for quantitative Real-Time PCR of the expression cloning DNA vector map to the Ori promoter region of Pfizer bivalent expression vector BNT162b2 (GenBank OR134577) and their sequences are published elsewhere.

Design and validation of quantitative multiplex Real-Time PCR

Since the declared mRNA in both Spikevax and BNT162b2 is modified with pseudouridine and it is not known to what extent this modification was performed by the manufacturer, it was not possible to have it artificially synthesized, as is a common practice for construction of calibration curves based on precise knowledge of the

sequence, sequence length, and mass of the synthesized target sequence.

Therefore, we used an alternative method where the Spikevax cDNA and BNT162b2 cDNA were diluted in serial log dilutions to assess whether the difference in the amplification curves for a given assay corresponds to a difference of approximately 3 cycles (Ct), which is a parameter of a correctly designed test and optimal reaction efficiency of all reaction components. If the measured difference in Ct values in individual logarithmic dilutions is approximately 3 cycles, i) the measurement results can be considered valid and ii) it is possible to use a universal calibration curve, which we routinely employ for similar purposes in cases when quantitation of an unknown target is needed.

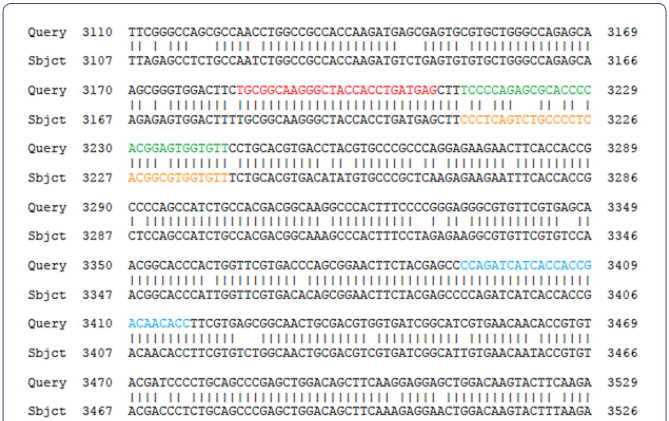


Figure 6: Region near the 3' end of the S protein mRNA of Spikevax (Query) and BNT162b2 (Subject) sequences targeted by quantitative Real-Time PCR. Common forward primer (red), common reverse primer (blue), probe-discriminating region (BNT162b2 in green - FAM, Spikevax in orange - ROX).

The universal calibration curve was created by averaging calibration curves constructed using serial dilutions of synthetic, precisely defined genomic fragments of 50 microorganisms, including ssRNA viruses, dsRNA viruses, ssDNA viruses, dsDNA viruses, bacteria and fungi (molecular targets that the laboratory routinely diagnostically examines using quantitative Real-Time PCR).

Oligonucleotide	Sequence in 5'-3'
Pfi_Mo common F	TGCGGCAAGGGCTACCACCTGATGAG
Pfi_Mo common R	GGTGTGTGCGGTGGTGATGATCTGG
VV_Ori_F	CTACATACCTCGCTCTGCTAATC
VV_Ori_R	GCGCCTTATCCGGTAACTATC
EcoliITS-F03	CACTCAGGCCTACCAAATTGCA
EcoliITS-R02	TCGAGTGAACCTTTGCAGGTAC
EcoliTSProbe_15	HEX - CGCATAGCTCCACCATCTCTGTAGTG - BHQ1
Spikevax Probe	ROX - TCCCCAGAGCGCACCCACGGAGTGGTGTT - BHQ2
VV_Ori_Probe	CY5 - TGCTGCCAGTGCGGATAAGTCGTGCTT - BHQ2
BNT162b2 Probe	FAM - CCCTCAGTCTGCCCTCACGGCGTGGTGTT - BHQ1

Table 1: Primes and fluorescent labelling.

The universal calibration curve equation we used in this investigation was

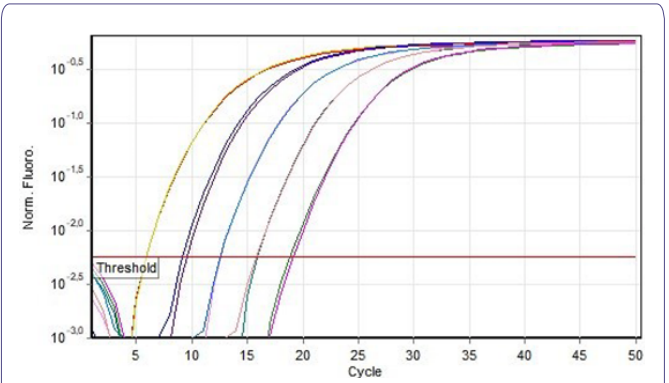
[Eq. 1] $\text{conc} = 10^{-(0.257 \cdot \text{Ct} + 11.897)}$.

Using this equation, we calculated the number of copies of the target sequence in a given volume using the obtained Ct values for each individual amplicon.

Quantitative Real-Time PCR was performed using FastStart™ Taq DNA Polymerase, Roche, USA, with 2 mM MgCl2 final. Nucleotides were purchased from Sigma, DE.

The temperature profile for amplification of all molecular targets in quantitative multiplex Real Time PCR was as follows: 94°C 5 min initial denaturation, then 50 cycles: 94°C for 20 sec, 57°C for 30 sec, then 72°C for 30 sec.

These results for Moderna and Pfizer are shown in figures 7 & 8 respectively.



No.	Color	Sample	Ct
1	Red	Spikevax 200100A - conc cDNA	5.90
2	Yellow	Spikevax 200100A - conc cDNA	5.91
3	Blue	Spikevax 200100A - 10x dil	9.12
4	Purple	Spikevax 200100A - 10x dil	9.50
5	Pink	Spikevax 200100A - 100x dil	12.59
6	Light Blue	Spikevax 200100A - 100x dil	12.53
7	Teal	Spikevax 200100A - 1000x dil	15.88
8	Light Red	Spikevax 200100A - 1000x dil	15.79
9	Green	Spikevax 200100A - 10000x dil	18.78
10	Magenta	Spikevax 200100A - 10000x dil	19.09

Figure 7: Spikevax cDNA dilution experiment – concentrated cDNA and 4 serial log dilutions.

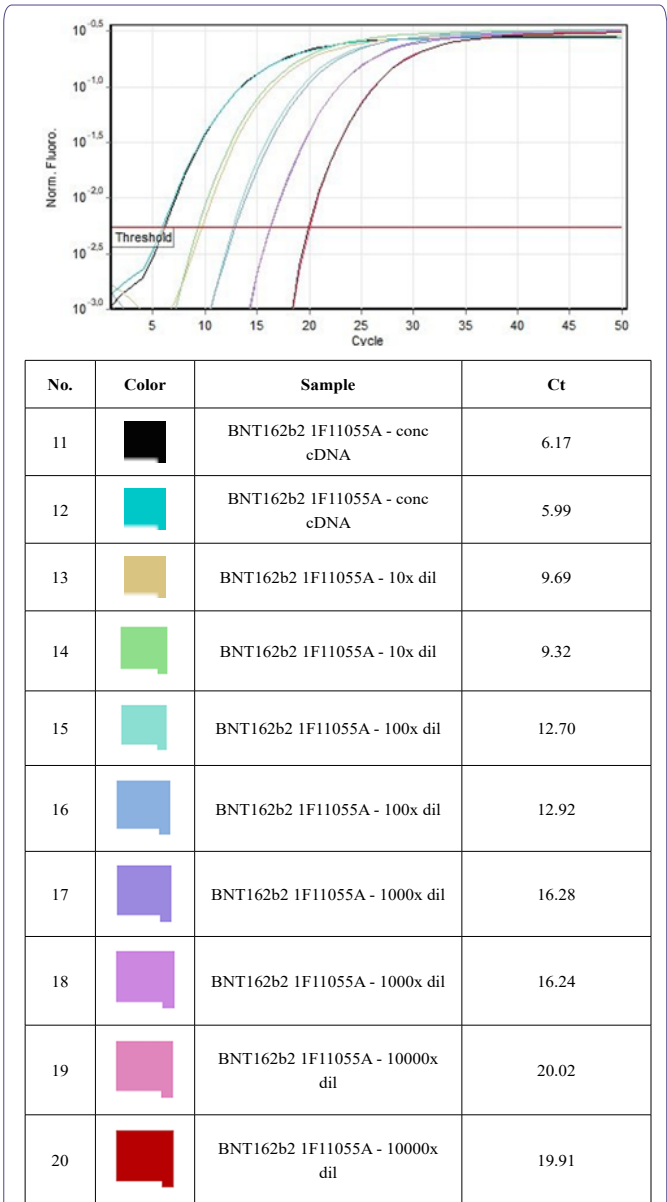


Figure 8: BNT162b2 cDNA dilution experiment - concentrated cDNA and 4 serial log dilutions.

Presence of undeclared DNA sequences

A notable observation made during this initial validation was that the cDNA isolates were heavily contaminated with the expression cloning vector.

These unexpected DNA sequences were identified in several vaccine lots. Analysis confirmed the presence of bacterial genomic DNA fragments from Escherichia coli, specifically sequences from the 16S rDNA internal transcribed spacer (ITS) region. These DNA elements are associated with bacterial vectors used during mRNA manufacturing.

For the sake of completeness, we carried out serial log dilution curves of the DNA of the contaminating expression vector, which were measured using the quantitative multiplex Real-Time PCR test used, essentially as an incidental finding against the background of

the primary goal - to validate the reaction efficiency of the detection of the manufacturer’s declared target, i.e. mRNA for the S protein.

These DNA amplicons also showed acceptable reaction efficiency, based on the measured Ct values in the dilution experiment as shown in figures 9 & 10.

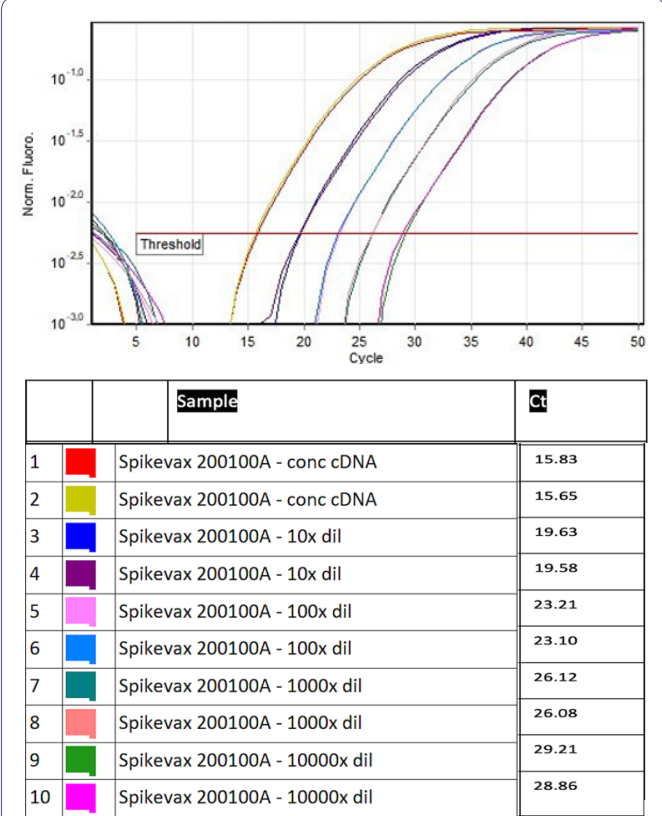


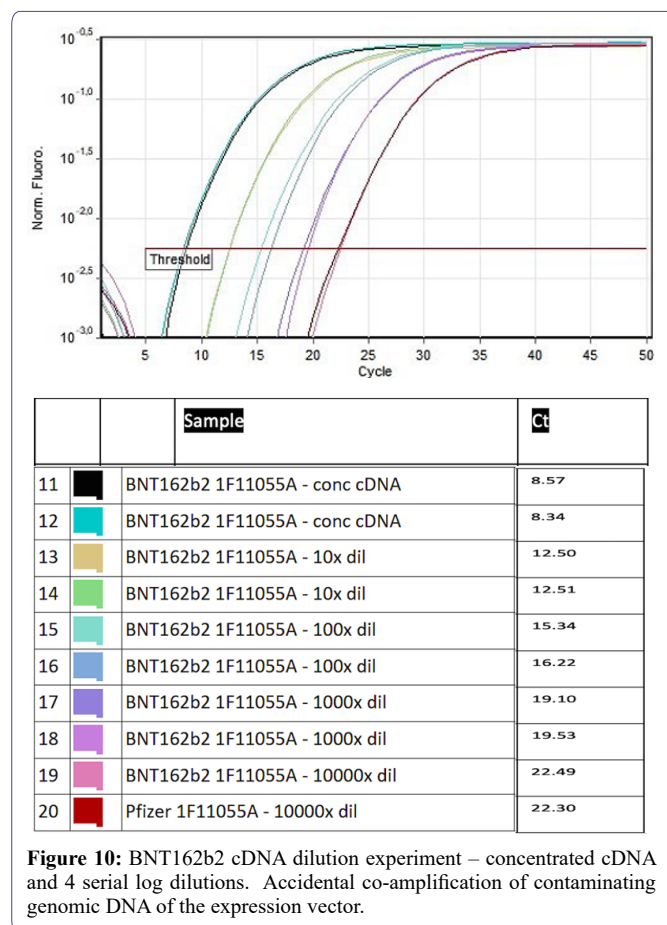
Figure 9: Spikevax cDNA dilution experiment – concentrated cDNA and 4 serial log dilutions. Accidental co-amplification of contaminating genomic DNA of the expression vector.

Oligonucleotides for quantitative Real-Time PCR of the Ori promoter expression cloning Sequencing of bivalent Moderna and Pfizer mRNA vaccines demonstrated nanogram to microgram quantities of expression vector dsDNA per dose. No evidence of SV40 was identified.

Oligonucleotides for quantitative Real-Time PCR of Escherichia coli genomic DNA were validated based on the requirements of ISO 13 485 as a constituent of a commercial diagnostic kit.

The oligonucleotides for quantitative Real-Time PCR cassettes for S protein (mRNA) in Spikevax and BNT162b2 preparations, newly used in this work, were verified by direct sequencing of obtained PCR products on an ABI 3500 capillary sequencer (ThermoFisher Scientific, USA).

All 17 lots of Spikevax and all 7 lots of BNT162b2 were sequenced. Direct sequencing of PCR products was performed using the BigDye™ Terminator v3.1 Cycle Sequencing Kit (ThermoFisher Scientific, USA). Sequencing reactions were purified using the BigDye XTerminator™ Purification Kit (ThermoFisher Scientific, USA), according to the manufacturer’s recommended procedure.



One typical example of a fragment of the chromatogram of the Spikevax S protein gene and one typical example of a fragment of the chromatogram of the BNT162b2 S protein gene are shown in figure 11. ABI 3500 sequencing data of all other lots are available upon request.

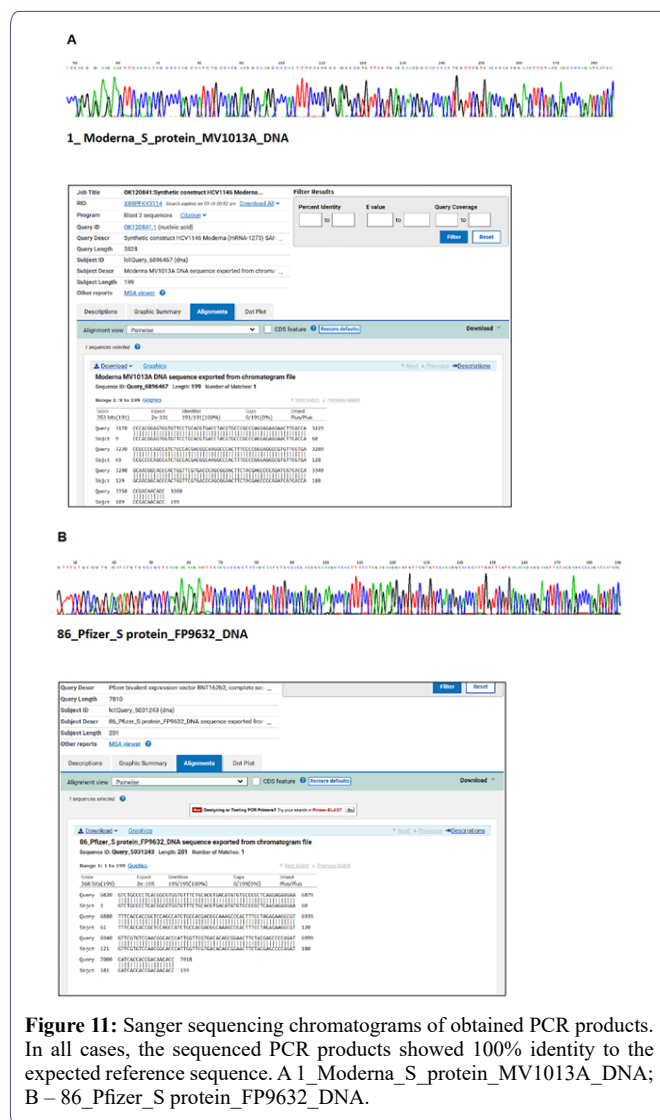
Quantitative real-time PCR analysis of mRNA for S protein

At the mRNA (cDNA) level, it was found that the expression of the declared molecular target (S protein mRNA) varies across lots for both Spikevax and BNT162b2.

For Spikevax (Moderna), as shown in figure 12, lots 200106A and MV1018A are characterized by one order of magnitude (10x) lower expression of S protein mRNA than the other tested lots. The blue dots in the graph indicate the S protein mRNA expression values for individual measurements (in the range of 10e9 to 10e10 copies of the target sequence/ml; analysis performed in quintuple for each lot). The orange dots in this case indicate the quantity of expression cloning vector DNA (in the range of 10e7 to 10e8 copies of the target sequence/ml; analysis performed in quintuple for each lot) present in the analyzed cDNAs. Similar findings are shown in figure 13, lots 1L084A and 1F1059A for Pfizer BNT162b2.

Quantitative Real-Time PCR analysis of DNA targets - S protein cassette and expression cloning vector promoter

A significant finding is the high quantity of DNA present across all tested lots, both Spikevax (Moderna) and BNT162b2 (Pfizer). In



all preparations, a very high Real Time PCR signal from the promoter of the cloning expression vector and from the 3' end of the cassette encoding the S protein was measured (10e7 – 10e9 copies/ml for Spikevax; 10e8 – 10e9 copies/ml for BNT162b2). Analysis of all lots was carried out in quintuples with the final results displayed in figures 14 & 15 respectively.

These data suggest that both assays might detect identical DNA construct, targeted at both its 5' end (promoter Ori) and its 3' end (bases 3184 to 3417 of the total 3880 bp of the complete coding sequence for the S protein). This most likely suggests that both Spikevax and BNT162b2 might contain the complete coding DNA sequence for the S protein cassette, together with some regulatory promoter sequences.

The results demonstrate more than degraded or fragmented DNA, that might arise from instability of the stored preparations or during suboptimal manufacturing process. The combined signal originates very likely from a full-length DNA construct, which might be capable of encoding a full-length mRNA for the S protein.

Within the tested lots of Spikevax (Moderna), the heterogeneity in the quantity of DNA coding for the S protein cassette and/or the

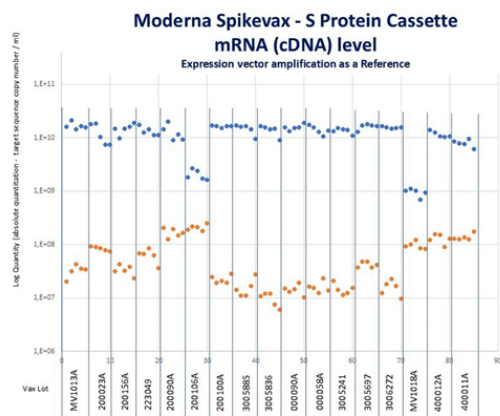


Figure 12: The graph shows the mRNA expression level for S protein in Spikevax (Moderna), GenBank OK120841 - blue dots in quintuplicates for each tested lot. Orange dots serve as a reference; it is an admixture of double-stranded DNA of the expression cloning vector in the tested cDNAs.

cloning expression vector is remarkable. In three cases, the ratio of the amount of expression cloning vector and DNA cassette for the S protein is reversed, and the difference in quantity is of one order of magnitude. This indicates that the given lot might also contain another construct cloned in the given expression vector. It is possible it could represent an admixture of Omicron, although this could mean that its amount is very random across the individual lots, which would raise concerns regarding good manufacturing practice.

We did not search for the Omicron sequence in this investigation. Rather, we focused on sequences that are clearly declared to be in the official content of the preparations – thus, we targeted the S protein mRNA [6], GenBank OK120841 (Spikevax, Moderna) [7]; GenBank OR134577 (BNT162b2, Pfizer) [8].

These differences in the quantity of DNA amplicons are clearly visible already in the raw data from the analyzer (RotorGene Q, Qiagen, Germany). Note the significantly different starts of the amplification curves.

Figure 16 shows the raw data analyzed at the DNA level for the S protein cassette Spikevax and BNT162b2, as well as for the expression cloning vector, which, like the DNA cassette for the S protein, is present in all tested lots.

The summarized results of the mRNA and DNA quantities for the S protein cassette are shown in table 2.

Quantitative detection of genomic DNA of escherichia coli

The presence of *Escherichia coli* genomic DNA was also analyzed within the multiplex quantitative Real-Time PCR to verify whether the preparations were contaminated with GMO, typically used for the propagation of the cloning expression vector harboring the S protein cassette.

A borderline quantity, in individual copies of the microorganism/ml of sample, was found in two lots of BNT162b2 (Pfizer), namely FP9632 and 1F1047A.

The data is not shown, though they can be provided upon request.

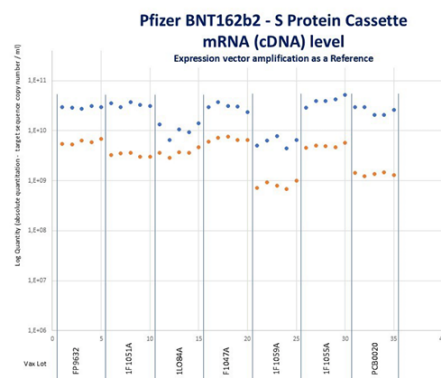


Figure 13: In BNT162b2 (Pfizer), the variation in mRNA expression for the declared S protein (GenBank OR134577) between individual lots is also noticeable – a difference of one order of magnitude (10x) is evident for lots 1L084A and 1F1059A – mRNA expression for S protein is shown in the graph by blue dots. The orange dots serve as a reference; it is an admixture of double-stranded DNA of the expression cloning vector in the tested cDNAs. Analysis of all lots was carried out in quintuples.

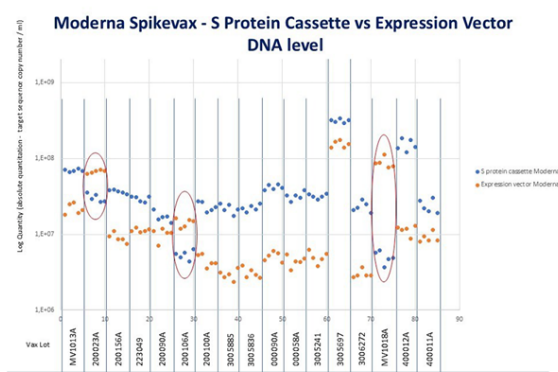


Figure 14: Shows a high quantity of DNA for both the expression cloning vector and the DNA for the S protein cassette. Of note are lots 200023A, 200106A, MV1018A, where the ratios of vector vs cassette are reversed. It is also interesting that lots 3005697 and 400012A are quantitatively significantly outside the range of the other tested lots.

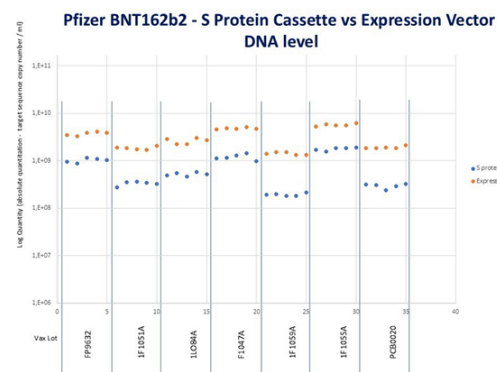
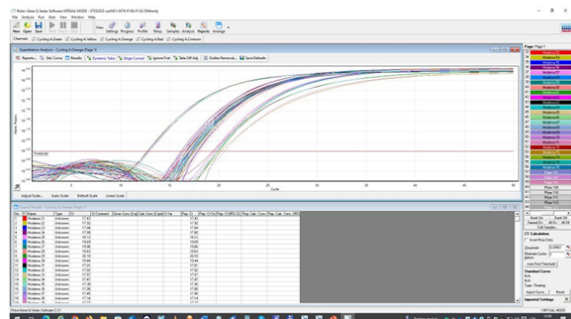
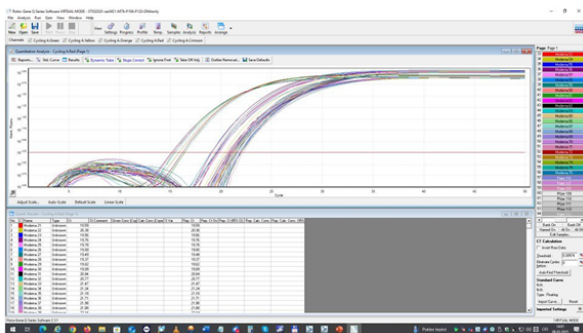


Figure 15: Shows a similar situation for BNT162b2 (Pfizer) preparations. In these preparations, the amount of expression cloning vector is higher, but what is more important is the horizontal heterogeneity within the blue dots, which represent quantitative S protein DNA cassette amplicons. Note the one order of magnitude (10x) difference between the lots.

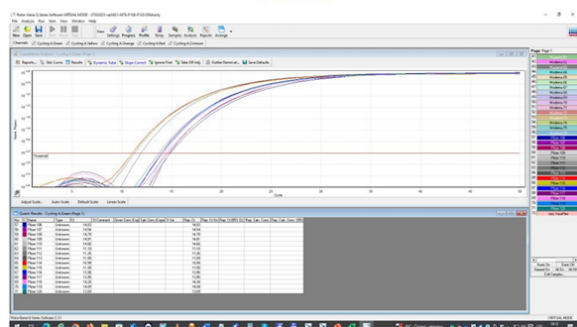
Moderna Spikevax - S Protein cassette - ROX DNA level



Moderna Spikevax - Expression Vector – Cy5 DNA level



Pfizer BNT162b2 - S Protein cassette - FAM DNA level



Pfizer BNT162b2 – Expression vector – Cy5 DNA level

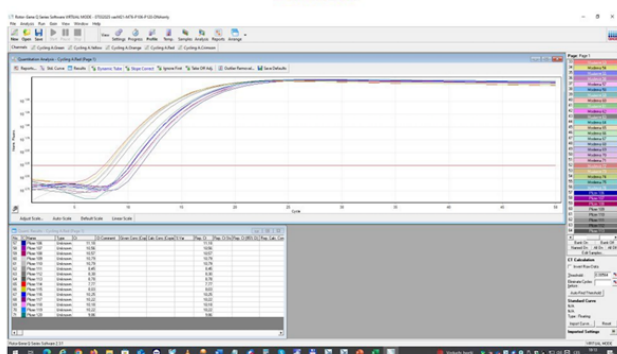


Figure 16: The following graphic print screens show raw data for the DNA cassette for the S protein (Spikevax - ROX, BNT162b2 – FAM, expression cloning vector – Cy5).

Homogeneity assessment

While there was a 28% batch-to-batch variability in nucleic acid content, there was no significant intra-lot inconsistencies observed. Each set of vials within a given lot showed consistent mRNA and DNA profiles.

Impact of expired storage conditions

Analysis of expired vaccine samples stored at -80°C indicated partial degradation of nucleic acid content. While mRNA fragments remained detectable, their integrity was reduced, potentially compromising vaccine effectiveness.

Quantities of S protein cassettes on mRNA (cDNA) and DNA levels in Moderna Spikevax and Pfizer BNT162b2 vaccines

	S protein cassette mRNA (cDNA) (copy number / ml)	S protein cassette DNA (copy number / ml)
Moderna Spikevax	$10e9 - 10e10$	$10e7 - 10e9$
Pfizer BNT162b2	$10e10 - 10e11$	$10e8 - 10e9$

Table 2: Summarized mRNA and DNA quantities in the Moderna and Pfizer Vials.

Discussion

This research study conducted Molecular analysis of the following lots of Moderna and Pfizer COVID genetic vaccines:

Spikevax (Moderna): MV1013A, 200023A, 200156A, 223049, 200090A, 200106A, 200100A, 3005885, 3005836, 000090A, 000058A, 3005241, 3005697, 3006272, MV1018A, 400012A, 400011A, and

BNT162b2 (Pfizer): FP9632, 1F1051A, 1L084A, 1F1047A, 1F1059A, 1F1055A, PCB0020.

Analysis of the samples using quantitative multiplex Real-Time PCR revealed:

- Inter-individual heterogeneity between individual lots is remarkable, with a difference in the quantity of declared mRNA of up to 10x.
- All Spikevax (Moderna) and BNT162b2 (Pfizer) lots tested contain a significant admixture of DNA, which is most likely the complete cassette for S protein cloned in an expression vector carrying intact regulatory sequences. Thus, expression of full-length mRNA for S protein from this DNA cassette cannot be excluded.
- The Real-Time PCR measured quantity of DNA in all preparations is comparable to the quantity of the only officially declared item, which is mRNA for S protein. This suggests that it is therefore not DNA “contamination”, but rather a regular admixture, undeclared by either manufacturer.
- Based on the Real-Time PCR measurement of the quantity of DNA present, another degree of heterogeneity can be observed between individual lots, where the possibility that other DNA construct(s), which identity is currently unknown, might also be present in some lots.

The presence of *Escherichia coli* DNA sequences raises questions about manufacturing quality control. While residual DNA from bacterial propagation systems is not uncommon in recombinant DNA products, its detection in these vaccines suggests incomplete purification during production. Regulatory standards such as those outlined by the European Medicines Agency (EMA) and the U.S. Food and Drug Administration (FDA) set limits on allowable residual DNA [4,5]. The observed DNA content may exceed these limits and pose theoretical risks related to genomic integration or immunological responses.

Furthermore, the observed variability in mRNA content may affect dose consistency. mRNA vaccines rely on precise nucleic acid delivery to ensure effective antigen expression. Deviations of 20–30% across lots, as observed in this study, may lead to inconsistent immune responses and altered clinical outcomes.

The continued use of the Wuhan strain's S protein sequence in both vaccines highlights a notable concern. Since early 2020, SARS-CoV-2 variants with significantly altered spike protein structures have dominated viral circulation. The use of an outdated sequence may reduce the vaccines' protective efficacy, particularly against newer variants with substantial antigenic drift.

The findings regarding mRNA degradation in expired samples emphasize the importance of strict adherence to storage protocols. Even with ultra-cold storage conditions, prolonged storage beyond designated expiration dates resulted in partial mRNA degradation, potentially impairing vaccine performance.

The above data shows a high degree of heterogeneity between individual Spikevax (Moderna) and BNT162b2 (Pfizer) preparations, measured by the quantity of declared mRNA for S protein. The individual lots differ in the quantity of the target mRNA sequence by 1 order of magnitude (10x).

In all preparations tested, a high quantity of double-stranded DNA (cloning expression vector, DNA cassette for S protein) was identified. Interestingly, the quantity of DNA was comparable to the quantity of mRNA declared by the manufacturer.

Such a high quantity of DNA clearly cannot be considered mere "contamination" during the manufacturing process. In the case of "a contamination", the quantity of the contaminating DNA would be expected many orders of magnitude lower, in the range of approximately 10e2 or 10e3 copies/ml. A random "contamination" of the Spikevax and BNT162b2 preparations with double-stranded DNA during the manufacturing process can therefore be excluded.

Moreover, unremarkable, borderline, contamination with genomic DNA of *Escherichia coli* (GMO used for large-scale production of cloning vectors) was found only in two cases of the BNT162b2 preparation (Pfizer), in individual units of copies of the target microorganism / ml. This might be considered a negligible finding in comparison to the large quantities of DNA of cloning vector and S cassette found in all lots tested. No SV40 was identified.

The presence of such high quantities of DNA in all the lots tested implies that it might not result from some contamination during an inadequate manufacturing process but rather might be considered a regular (though not officially declared) constituent of all lots tested, present in quantities (almost) identical to the quantity of mRNA for the S protein.

Both Pfizer and Moderna identified the only oligonucleotide materials within their vaccines as being mRNA. The presence of double-stranded DNA, or any other DNA, in the Moderna and Pfizer preparations, was not declared by either the manufacturers.

Conclusion

This study highlights key findings regarding the oligonucleotide content of Spikevax (Moderna) and BNT162b2 (Pfizer) COVID-19 vaccines. The presence of these genetic sequences also raises InflammoThrombotic Immunologic Response (ITIR) concerns [1]. While declared mRNA sequences were confirmed, variability in nucleic acid content and the presence of undeclared DNA sequences underscore the need for improved quality control in vaccine manufacturing.

Key recommendations include:

- **Enhanced Purification Protocols:** Manufacturers should review DNA removal processes to minimize residual contamination and reduce InflammoThrombotic Immunologic Response (ITIR).
- **Stricter Lot Testing:** Additional oversight is recommended to ensure batch-to-batch consistency in nucleic acid content.
- **Genomic Updates:** Given the ongoing evolution of SARS-CoV-2 variants, vaccine designs should align with contemporary circulating strains.
- **Removal of contaminated genetic vaccines from the market.**
- **Stricter oversight by regulatory agencies including the FDA, EMA and ŠÚKL.**

Continued vigilance in vaccine production and regulatory oversight is crucial to ensuring public confidence in mRNA and DNA vaccine technology and maximizing global immunization efforts.

Funding

This study was carried out in response to a formal request by Slovakian Prime Minister Mr. Robert Fico in and through the office of the Plenipotentiary for the Slovak Republic to assess the nucleic acid content in multiple lots of Spikevax and BNT162b2 vaccines.

IRB: No Institutional Review Board required.

Conflict of Interest

The authors declare no conflict-of-interest.

Data Availability

Further data is available if determined appropriate by submission request including but not limited to the individual(s) and institution(s), purpose/intent of request, and pertinent relevant requested information as deemed applicable by the authors. <https://www.flemingmethod.com/>

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